



The D614G Virus Mutation Enhances Anosmia in COVID-19 Patients: Evidence from a Systematic Review and Meta-analysis of Studies from South Asia

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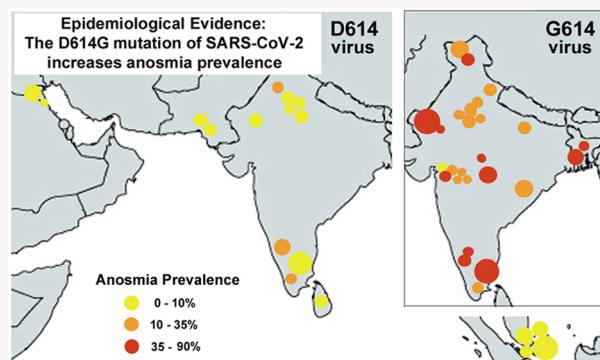
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ABSTRACT: The prevalence of chemosensory dysfunction in patients with COVID-19 varies greatly between populations. It is unclear whether such differences are due to factors at the level of the human host, or at the level of the coronavirus, or both. At the host level, the entry proteins which allow virus binding and entry have variants with distinct properties, and the frequency of such variants differs between ethnicities. At the level of the virus, the D614G mutation enhances virus entry to the host cell. Since the two virus strains (D614 and G614) coexisted in the first six months of the pandemic in most populations, it has been difficult to distinguish between contributions of the virus and contributions of the host for anosmia. To answer this question, we conducted a systematic review and meta-analysis of studies in South Asian populations when either the D614 or the G614 virus was dominant. We show that populations infected predominantly with the G614 virus had a much higher prevalence of anosmia (pooled prevalence of 31.8%) compared with the same ethnic populations infected mostly with the D614 virus strain (pooled anosmia prevalence of 5.3%). We conclude that the D614G mutation is a major contributing factor that increases the prevalence of anosmia in COVID-19, and that this enhanced effect on olfaction constitutes a previously unrecognized phenotype of the D614G mutation. The new virus strains that have additional mutations on the background of the D614G mutation can be expected to cause a similarly increased prevalence of chemosensory dysfunctions.

KEYWORDS: COVID-19, anosmia prevalence, olfactory dysfunction, SARS-CoV-2, D614G virus mutation, South Asia



INTRODUCTION

Chemosensory dysfunction has been identified as one of the most frequent symptoms of COVID-19.^{1–4} However, the prevalence of smell loss in COVID-19 varies greatly between populations. Some studies reported the prevalence of anosmia or hyposmia to be lower than 1%, while others reported a prevalence of over 70%.^{1,2,4–7} Such differences may be due to multiple factors, but two reasons are currently considered to be most relevant: differences at the level of the human host (variants in the virus entry proteins, angiotensin-converting enzyme 2, ACE2, or in the protease TMPRSS2) and differences at the level of the virus (mutations of the spike protein resulting in an altered efficiency of entry to the host cell and, therefore, higher infectivity), or a combination of these host and virus factors.^{1,2,4,5,8–13}

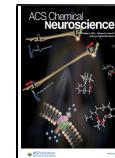
For historical reasons of how the pandemic unfolded, it has been difficult to define the relative contributions of host and virus factors for anosmia. The pandemic started in East Asia with the less infectious D614 virus strain, but when COVID-19 reached Europe and North America and the rest of the world, the virus with a D614G mutation in the spike protein rapidly

replaced the original D614 virus.¹⁴ Since the pandemic was largely controlled in East Asia in the second half of 2020, there are very few studies from East Asia on prevalence of anosmia after the initial D614 virus infestation. Studies reporting anosmia prevalence from the rest of the world typically collected data from regions where the two virus strains coexisted or when the G614 virus was dominant.^{1,4,11,14} This made it impossible to discern whether the virus mutation or host protein variants are primarily responsible for the differences in anosmia prevalence between populations in East Asia and the rest of the world.

Here, we took advantage of the fact that early studies from South Asia collected data on anosmia prevalence in regions

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with 50% or more D614 virus infections, while more recent studies collected data in South Asia when the G614 strain was dominant. Our systematic review and meta-analysis of the same ethnicity (South Asians) shows a strong association between the D614 virus and low prevalence of anosmia and a strong association between the G614 virus and high anosmia prevalence. The most parsimonious interpretation of these data is that the D614G mutation plays a significant, and apparently the most important, role in causing an increased prevalence of anosmia in COVID-19 patients. We, here, present our evidence, and we discuss the presumed underlying molecular mechanisms of how SARS-CoV-2, and especially the G614 virus, causes olfactory dysfunction.

Spread and Increasing Dominance of the G614 Virus over the D614 Virus. In the beginning of the pandemic, the D614 virus emerged in China and spread to the rest of Asia, Europe, the Middle East, Australia, and North and South America. The spread beyond East Asia was accompanied by the appearance of the more infectious G614 virus mutation, and over the next weeks and months this strain became dominant over the original D614 strain in most parts of the world.^{11,14,15} The D614 virus remained dominant beyond May of 2020 in only few regions: most of China, Singapore, Malaysia, the South of India, and the Delhi region (<https://www.gisaid.org/>; <https://cov.lanl.gov/apps/covid-19/map/>)^{14,16–25} The coexistence of the D614 and G614 viruses made it impossible, in most regions, to discern whether differences in the prevalence of anosmia were due to differences among the host populations (frequency of variants of the entry proteins ACE2 or TMPRSS2 and thus ethnic differences) or were due to differences in the infectivity and cell entry efficiencies of the coronavirus. In East Asia, nearly all studies were conducted when the D614 virus was dominant. In Europe, the Americas, the Middle East, and Africa, there was either a coexistence of D614 and G614 viruses, or a dominance of the G614 virus during the periods of data collection for anosmia prevalence. To clarify the suspected role of the virus type for anosmia, it was necessary to find an ethnicity which was infected either at different times or in different regions when either the D614 or G614 virus was dominant and to compare the prevalence of anosmia between studies reporting on such cohorts. There are no suitable and unambiguous (unequivocal) pairings of appropriate studies from Europe, the Middle East, the Americas, or East Asia that allow comparison of the same ethnicity for the two types of virus strains. Fortunately, we found that one ethnicity does meet these criteria—South Asians.

RESULTS

Spatiotemporal Mapping of D614 or G614 Virus Dominance in South Asians. The spread of the SARS-CoV-2 virus into the Indian subcontinent began at the end of January 2020, with waves from several regions: One from South East Asia (and the Middle East), carrying the D614 virus, and another wave primarily from Europe and the USA, carrying the G614 virus. Overall in India, the D614 virus dominated until late April or May, while the G614 virus became dominant in most Indian states mid to late May or June 2020.^{16,17,19–21,27,28} However, there were significant regional differences. For example, the D614 virus persisted for a longer time in the Delhi region^{17,19} and also in the South of India,^{17,20,21,24,27–30} while the G614 virus dominated early in the West of India (Gujarat, Maharashtra),^{17,19,25,31} and also

in the Eastern and Central Indian states,²⁸ as well as in Bangladesh.^{14,26,32,33} Pakistan and Sri Lanka had different waves of virus spread, with an early D614 dominance, but becoming all G614 in June and July 2020.^{34–36} Importantly, the predominance of the G614 virus was delayed in South India until mid May to June of 2020.^{17,18,20–22,25,30} Such regional differences were caused by travel routes, as well as regional spreader events, for example, in Delhi.¹⁷ Indians and Bangladeshis were exposed primarily to the D614 virus in March through June or July of 2020 in some regions outside the Indian subcontinent, for example, Kuwait,³⁷ Malaysia,^{38,39} and Singapore,^{14,23} while South Asians in Oman were mostly exposed to the G614 virus in March and April of 2020.^{37,40} The temporospatial distribution of the D614 virus versus G614 virus in South Asia is summarized in Figure 1A–F.

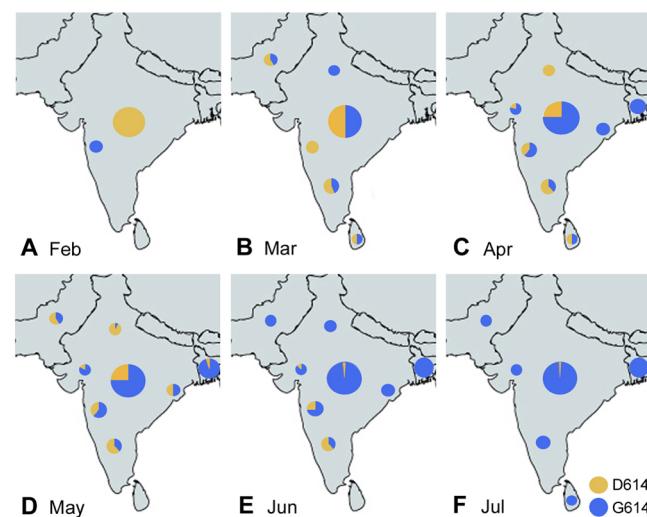


Figure 1. Map of South Asia showing contributions of D614 and G614 virus to COVID-19 in February to July of the year 2020. Virus strain dominance according to relevant references.^{14,16–22,24–29,31–36} The large circle in the center of India shows the overall contributions of D614 and G614, regardless of region; the small circles represent regional contributions of D614 and G614 during the indicated months of 2020.

Systematic Review and Meta-analysis of South Asian Studies. We, here, explore the hypothesis that there was a lower anosmia prevalence when the D614 virus was the prevailing strain, while a higher anosmia prevalence was induced by the G614 virus. Therefore, we searched the literature and conducted a systematic review and meta-analysis on studies reporting olfactory dysfunction in South Asian patients with COVID-19, sorted by dominance of the two virus strains. Dominance of D614 vs G614 virus was determined by reviewing studies that mapped the temporospatial changes within different regions of the Indian subcontinent.^{17,19,25,28,33–35} For our systematic review and search strategy, we adhered to the PRISMA guidelines⁴¹ (Figure 2).

Our search retrieved 598 studies that examined South Asians for COVID-related loss of smell, of which 40 met the inclusion criteria (see Methods). These 40 studies (reporting on 43 cohorts) were subjected to a meta-analysis (Table 1). Fifteen studies reported anosmia prevalence in 18 cohorts with a total of 7247 COVID-19 patients from regions where the D614 virus was dominant: three studies from Kuwait or Singapore^{42–44} and 12 from India or Pakistan.^{45–56} We compared

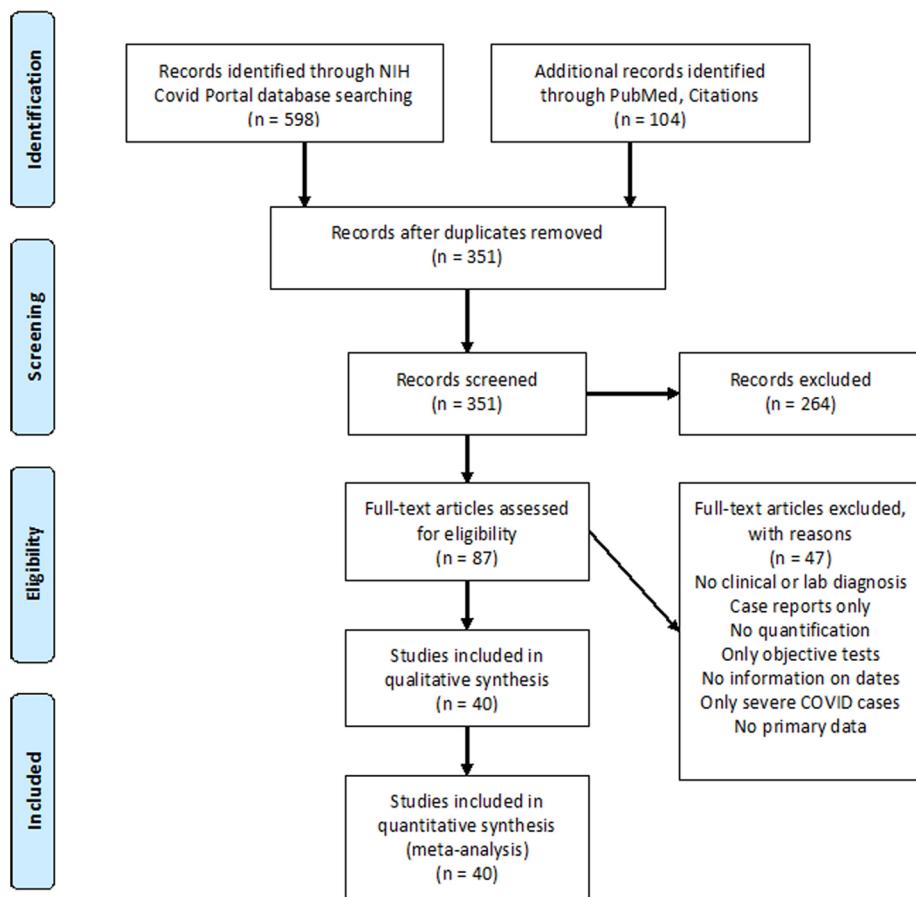


Figure 2. Flowchart of literature search and systematic review of studies reporting on COVID-19 related olfactory dysfunction in South Asians through July 22, 2021.

such data with the results obtained in 25 studies reporting on 25 cohorts with a total of 9626 South Asian patients from the Indian subcontinent (India and Bangladesh) and Oman, when the G614 virus had become dominant (Table 1, illustrated in Figure 3A and B).^{6,57–80} The differences in results between the two types of cohorts are shown in the forest plots (Figure 4A). The pooled prevalence of olfactory dysfunction in the same ethnicity (South Asians) in regions with D614 predominance was 5.33% (95% confidence interval, CI = 3.52–8.00%), while in regions with G614 predominance, it was 31.79% (95% CI = 23.26–41.76%) (Figure 4A, C). The subgroup test from the random effects meta-analysis showed that this was a statistically significant difference with $p < 0.0001$ (Figure 4C).

The main novel finding of our meta-analysis is that, when the same ethnicity is compared for anosmia prevalence under two different conditions, with either the D614 or G614 virus strain being dominant, there is a large difference in anosmia prevalence between the two conditions. This means that the more infectious virus type, G614, is a significant factor for anosmia prevalence, apparently more important than ethnic differences in variants of the host proteins, ACE2, and TMPRSS2. This is presumably due to a higher efficiency of G614 virus entry to the host cell,^{11,13,81} as discussed at the end of this Review.

EFFECTS OF RESPONSE BIAS, AGE, DISEASE SEVERITY, GENDER, AND METHODOLOGY

Are there alternative interpretations of our data that could explain, entirely or partially, the differences in anosmia prevalence among South Asian populations? Three types of parameters need to be considered: response bias, demographics, and methodology. If the demographics of the two cohort types (D614 vs G614 virus dominance) differed, they could have influenced or biased the comparisons. Demographics that have been shown or suspected in previous reviews to have an association with anosmia prevalence include age, gender, and COVID-19 disease severity. Age was associated in some analyses.^{1,82} Gender was found to show trends in the largest meta-analysis¹ but not in two earlier analyses that considered fewer studies.^{82,83} Multiple studies and reviews agree that disease severity is negatively associated with anosmia prevalence.^{1,83–86} Finally, objective methodology to assess olfactory dysfunction may be more sensitive than subjective recall, resulting in an increased prevalence, although studies are controversial.⁷ Response bias may occur if early studies did not inquire about olfactory dysfunction as much as later studies.

Response Bias. Early in the pandemic, there was no publicity about COVID-19 causing loss of smell. Patients, caregivers, and investigators may not have asked about this symptom or regarded it as irrelevant, resulting in response bias compared with later studies. This issue has been widely discussed.^{1,2,4,5,10,83,87,88} Could this bias explain why early

Table 1. Smell Dysfunction in COVID-19: Chronology of Studies on South Asians with Either D614 Virus Dominance or G614 Virus Dominance^a

author	ref	start data collection (mm/dd/yy or month and year)	end data collection (mm/dd/yy or month and year)	ethnicity (location) north, west, south, east India (N, W, S, E)	cohort no.	smell loss prevalence (%)	age (mean, m; median, M)	male gender (%)	severe COVID disease (%)
D614 virus dominance									
Almazedi	42	02/24/20	04/20/20	Indian (Kuwait)	527	0.1	M 41	81	1.1
Almazedi	42	02/24/20	04/20/20	Bangladeshi (Kuwait)	70	0.1	M 41	81	5.7
Varghese	45	Feb 2020	03 31 20	S India (Kerala)	202	12	m 36.5	80.2	3
Ish	46	03/01/20	04/30/20	N India (Delhi)	170	4.1	ND	ND	0
Sharma	47	03/20/20	04/30/20	N India (Rajasthan)	234	0.0	m 35.1	64.5	0
Tham	43	03/24/20	04/16/20	Bangladeshi (Singapore)	568	7.74	m 34	87.6	ND
Tham	43	03/24/20	04/16/20	Indian (Singapore)	221	9.95	m 34	87.6	ND
Herath	48	03/10/20	05/30/20	Sri Lanka	431	2.3	m 37	78	1.2
Iltaf	49	Mar 2020	Jun 2020	Pakistan (Karachi)	350	1.4	m 49.5	70	ND
Lal	50	03/01/20	06/30/20	N India (Delhi)	435	8.0	m 38.3	66.7	ND
Yadav	51	03/01/20	07/05/20	N India (Punjab)	152	18.4	m 43	51.3	0
Khurana	52	Apr 2020	Apr 2020	N India (Delhi)	94	3.2	m 36	59.6	1
Lanjiwar	53	04/01/20	04/20/20	N India (Delhi)	109	9.2	m 38.2	86.2	0
Krishnasamy	54	Apr 2020	May 2020	S India (Tamil Nadu)	1263	9.4	M 35	66.3	10.5
Smitha	55	Apr 2020	Jun 2020	S India (Karnataka)	500	13.4	~40	62	5
Makda	56	Apr 2020	Jul 2020	Pakistan (Karachi)	114	7.8	m 51	54.4	54.4
Soh	44	05/01/20	07/01/20	Indian (Singapore)	743	2.6	m 35	100	0
Soh	44	05/01/20	07/01/20	Bangladeshi (Singapore)	1064	4.7	m 35	100	0
total					6247	5.3% (mean)	38.83 (mean)	75.08 (mean)	5.85% (mean)
G614 virus dominance									
Al Harthi	57	03/03/20	05/09/20	South Asian (Oman)	102	6.9	m 26	77.5	18.6
Kuchhal	58	04/01/20	07/31/20	N India (Uttarakhand)	465	18.9	ND	ND	ND
Mishra	59	Apr 2020	May 2020	W India (Maharashtra)	74	14.8	~35	58.1	ND
Bhatta	60	04/01/20	07/30/20	W India (Maharashtra)	600	63.7	m 42.9	56.2	13.3
Panda	61	4/23/20	06/29/20	N India (Delhi)	225	22.1	m 35	70.7	0
Chaurasia	62	04/01/20	07/31/20	N India (Uttarakhand)	465	18.9	ND	ND	ND
Malik	63	04/29/20	07/20/20	Bangladesh	139	38.9	m 34.1	41	1
Shah	64	Apr 2020	Aug 2020	N India (Srinagar)	655	18.5	m 32.7	63.2	0
Bhattacharjee	65	05/12/20	05/21/20	W India (Maharashtra)	34	15	m 35	61.8	0
Jain	66	May 2020	Jun 2020	N India (Delhi)	410	22.4	m 37.2	63.9	0
Dev	67	05/01/20	06/15/20	N India (Delhi)	261	21	m 36	58	0
Kumar L	68	May 2020	Aug 2020	N India (Delhi)	141	24.1	m 15.2	58.9	0
Koul	69	05/15/20	08/15/20	N India (Jammu)	300	42.5	m 37	74	0
Sahoo	70	06/25/20	07/24/20	E India (Odisha)	718	10.7	m 34	78	0
Rajkumar	71	07/06/20	07/12/20	S India (Tamil Nadu)	230	26.9	m 43.6	63	0
Kumar V	72	Jun 2020	Aug 2020	N India (Delhi)	200	34.0	m 37.4	75.5	0
Kandakure	73	Jun 2020	Sep 2020	W India (Maharashtra)	200	8.5	~35	54.5	0
Hasan	74	Jul 2020	Sep 2020	Bangladesh	600	38.7	m 50.2	64	0
Hajare	75	Jul 2020	Sep 2020	S India (Karnataka)	167	62.9	m 39	54.5	ND
Thakur	6	Sep 2020	Oct 2020	S India (Karnataka)	250	71.6	~44	57.6	0
Savtale	76	10/01/20	10/15/20	W India (Maharashtra)	180	55.5	m 37.8	33.4	0
Gupta	77	Nov 2020	Nov 2020	N India (Rajasthan)	98	86.5	m 41.4	49	0
Mangal	78	11/16/20	11/26/20	W India (Maharashtra)	57	17.7	~35	ND	0
Karthikeyan	79	Aug 2020	Dec 2020	S India (Tamil Nadu)	1000	74.2	m 42.4	59.6*	0
Silu	80	Sep 2020	01/15/21	N India (Rajasthan)	2055	43.0	m 50.4	77.4	2.8
total					9626	31.8% (mean)	34.32 (mean)	61.35% (mean)	1.64% (mean)

^aAbbreviations: ND, not disclosed; *, 3.2% transgender was assigned 50/50 to male and female gender. Virus dominance was determined by multiple studies as described in the Spatiotemporal Mapping of D614 or G614 Virus Dominance in South Asians section.

studies in Asia did not report olfactory dysfunction as often as subsequent studies on Caucasians? This is unlikely to be a decisive factor for two reasons. Careful analyses of East Asians for chemosensory dysfunction revealed low prevalence,^{5,89,90} even with objective olfactory testing,⁹¹ and when the olfactory dysfunction was re-examined in Chinese populations at a later date, the anosmia prevalence still was significantly lower in Chinese patients than in Caucasian patients.⁹² Second, we

show here for South Asians that studies conducted later in the pandemic, when the media had widely publicized and revealed anosmia/hyposmia as a cardinal symptom of COVID-19, reported a low prevalence when they examined cohorts in a region (e.g., Delhi and Karnataka) and at a time when the D614 virus strain was dominant. Therefore, we can exclude response bias as the main explanation for prevalence differences in South Asians.

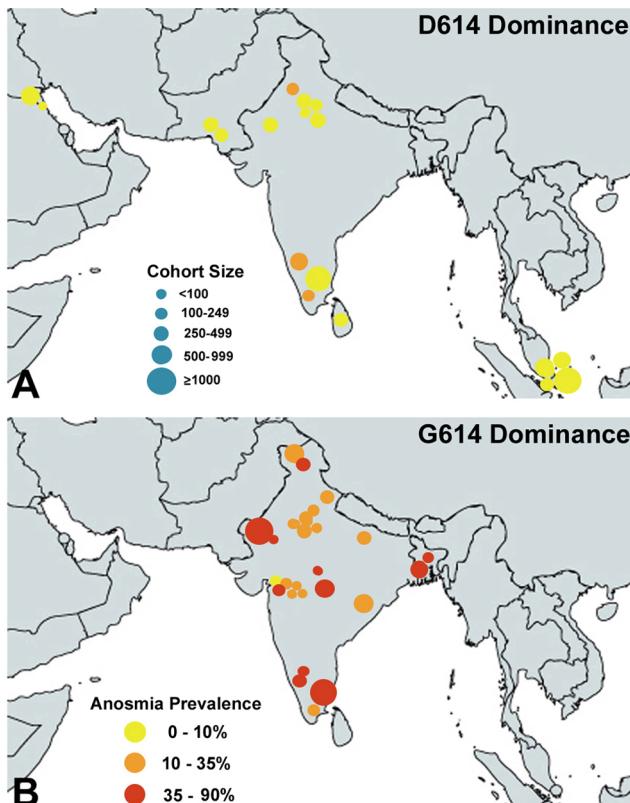


Figure 3. Location of studies reporting the prevalence of olfactory dysfunction among South Asians with D614 virus predominance (A) and G614 virus predominance (B). The cohort size is indicated by the size of the blue dots, the prevalence of olfactory dysfunction is indicated by the heat map, increasing from yellow to red. Note that mostly D614 infections lead rarely to a more than 10% anosmia prevalence, while almost all of the mostly G614 infections lead to a prevalence of 10–90% in the same ethnicity (South Asians).

Age. When we tested the parameter of age in our pooled analysis, we found no significant difference between the two groups (one group infected predominantly with the D614 virus, the other with the G614 virus). Mean age was $38.83 \text{ years} \pm 1.23 \text{ standard error (SE)}$ for the D614 group and $34.32 \text{ years} \pm 1.53 \text{ SE}$ for the G614 group, with $p = 0.3288$ ($\beta = 0.037 \pm 0.038 \text{ SE}$), which is no significant difference.

Disease Severity. As a measure of COVID disease severity, we calculated the percentage of hospitalizations or percentage of cases with “severe” disease, when disclosed, and we found in our pooled analysis that the D614 group had a mean 5.85% severe cases $\pm 3.82 \text{ SE}$, while the G614 group had $1.64\% \pm 1.05 \text{ SE}$. The subgroup test showed no significant difference in anosmia based on disease severity with $p = 0.5294$ ($\beta = -0.0173 \pm 0.027 \text{ SE}$). We conclude that disease severity does not explain the prevalence differences between the cohorts with D614 or G614 virus dominance.

Gender. It is unclear whether gender is associated with anosmia in COVID.^{1,10,82,83} We tested the parameter of gender using percent male in each cohort as a continuous variable in a meta-regression. In both groups, there were more males than females. Specifically, among the D614 group, there was a mean of $75.08 \pm 3.57\%$ males, while there was a mean $61.35 \pm 2.45\%$ males in the G614 group (mean \pm SE). The subgroup test was significant ($p < 0.0001$) with a negative linear relationship between percent of cohorts that were male and the

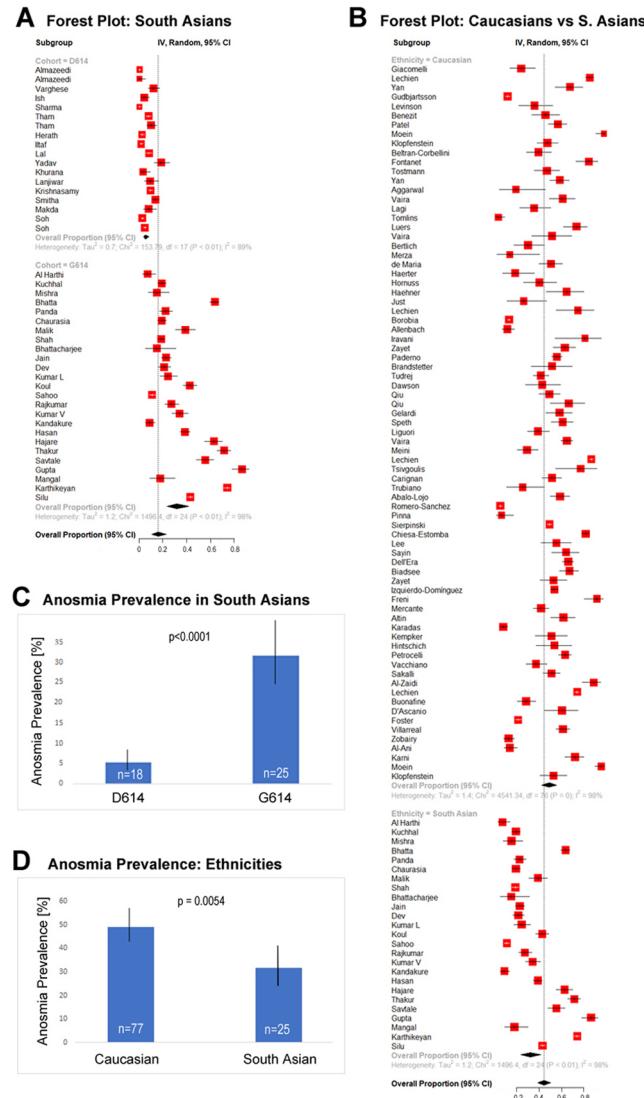


Figure 4. Comparison of the prevalence of olfactory dysfunction in populations infected with D614 or G614 virus predominance. (A) Forest plots of olfactory dysfunction prevalence in South Asians infected mostly with D614 vs G614 virus. (B) Forest plots of olfactory dysfunction prevalence in Caucasians vs South Asians infected mostly with the G614 virus. (C) Bar graph comparing the pooled anosmia prevalence with 95% confidence intervals and p -value between D614 and G614 cohorts. (D) Bar graph showing the pooled prevalence of anosmia in Caucasians vs mostly G614 virus-infected South Asians (Caucasian cohorts from von Bartheld et al., 2020).¹ Numbers of cohorts are indicated in white on the bars.

prevalence of anosmia ($\beta = -0.0586 \pm 0.0144 \text{ SE}$; Supporting Figure 1). Accordingly, a contributing effect of gender cannot be ruled out. However, the possible gender effect on anosmia prevalence is small: we calculated that the difference in the male/female ratio between the two cohort groups (Table 1) would account for less than 1/10th of the observed difference in anosmia prevalence between the two groups (5.3% vs 31.8%, Figure 3A).

Multivariable Meta-regression. When we modeled age, gender, and group (D614 or G614 virus dominance cohorts) together, cohort remained highly significant at $p < 0.0001$ ($\beta = 1.95 \pm 0.363$, D614 = reference), while gender and age showed marginal significance at $p = 0.0549$ and $p = 0.0484$, respectively

($\beta_{\text{gender}} = -0.0235 \pm 0.0122$, $\beta_{\text{age}} = 0.0477 \pm 0.0242$). We did not include disease severity in our multivariable meta-regression due to missing data: 76.7% ($n/N = 33/43$) of the cohorts provided data on all four variables, whereas modeling only age, gender, and group allowed us to use 90.70% of the cohorts ($n/N = 39/43$) in our multivariable regression.

Methodology. Chemosensory dysfunction in COVID-19 can be assessed by questionnaires and history taking (subjective tests) or by testing the sense of smell objectively.^{1,7,93} Some studies using objective tests showed an increase in the prevalence of olfactory dysfunction,^{1,2,82,93–95} while other studies reported the opposite (reviewed in Boscutti et al., 2021).⁷ In the studies on South Asian COVID-19 patients, objective tests were used in only three of the cohorts with G614 virus dominance. To avoid methodology as a confounding variable, we considered only studies with subjective questioning. We can, therefore, rule out methodological parameters as an explanation for the difference in anosmia prevalence between D614 and G614 cohorts.

Taken together, we conclude that response bias, age, disease severity, and methodology cannot sufficiently explain the difference in prevalence between the two types of cohorts, and that after adjusting for the effects of age and cohort virus type, the effect of gender is relatively small compared to that of the cohort virus effect. Accordingly, given that we controlled for ethnicity by including only cohorts with South Asian populations, the virus type (G614 vs D614) appears to be the most relevant parameter that is responsible for the vast majority of the observed prevalence differences.

LIMITATIONS

Sampling of virus genomics differs between regions in South Asia, some regions have low sample numbers, and results may not be representative for all of these regions. Some studies reported only approximate dates for the beginning and end dates of study periods; furthermore, when data on anosmia were collected over a longer period of months, it is possible that the majority of patients were enrolled early or late during that period, making the correlation with the virus strain less precise. We cannot entirely rule out a small contribution of response bias—earlier studies may have insisted less on information about olfactory dysfunction than later studies. Finally, there is some heterogeneity, possibly due to yet unknown parameters and lack of information about disease severity among patients within cohorts.

SUPPORTING EVIDENCE FOR A ROLE OF G614 IN ANOSMIA PREVALENCE

Our meta-analysis of studies on South Asians provides a strong case for a role of the D614G virus mutation in anosmia prevalence. Is there additional evidence from studies on other ethnicities to support this notion? The study by Eyre et al. (2020)⁹⁶ examined anosmia prevalence in the UK separately for Caucasians, Asians, and Chinese subjects residing in the UK and found no significant difference in prevalence between ethnicities at a time when the G614 virus was predominant. This supports the idea that ethnicity plays no or only a minor role in anosmia prevalence, although the data on “Chinese” was based on a very small cohort ($n = 10$).

The prevalence of anosmia was also examined by three studies in Hong Kong,^{97–99} at a time when the G614 virus began to dominate over the D614 virus (March–April

2020).^{14,100} Apparently, the G614 virus was dominant in Hong Kong because of the extensive travel to Hong Kong from the UK and USA,⁹⁹ where the G614 virus had taken over. This region is the one exception to the predominance of the D614 virus throughout the rest of mainland China in 2020.¹⁴ All three of the studies in Hong Kong found the prevalence of anosmia to be much higher (66.7%, 47.0%, and 22.1%, – weighted mean = 37.44%) than the pooled prevalence estimated for other regions in East Asia (16.7%) using meta-analytic techniques,¹ although one of the studies from Hong Kong reported on a very small cohort ($n = 18$).⁹⁷ The studies from Hong Kong again point to the G614 virus as the reason for the increased anosmia prevalence. When anosmia prevalence was determined early during the pandemic in Malaysia (in April 2020), while the D614 virus was still dominant, with only a minor contribution of the G614 virus, the anosmia prevalence was 21.4%.³⁷ Just 3–5 months later, when the G614 virus dominated,^{14,38} the anosmia prevalence had increased to 36.6%,¹⁰¹ consistent with our hypothesis.

There are some other indications, from regions in Europe, that a larger fraction of the D614 virus early in the pandemic may have contributed to a relatively low anosmia prevalence, for example, in Iceland¹⁰² and in Spain.¹⁰³ However, while suggestive, the period of data collection in these studies was at a time when the two virus strains coexisted, making it impossible to know whether the patients with low anosmia prevalence were indeed mostly those infected with the D614 virus. Furthermore, the study from Spain¹⁰³ primarily examined hospitalized patients, and disease severity associates negatively with anosmia prevalence, as mentioned above. Taken together, the Hong Kong studies^{97–99} provide the most convincing supporting evidence outside of South Asia for a contribution of the G614 virus to anosmia.

DISCUSSION

Is There Also a Contribution of Human Host Variants—True Ethnic Differences? There are several variant proteins in the human host that could cause ethnic differences in anosmia prevalence. The one that has been most discussed is ACE2, which has variants that are known to differ in their binding affinities to the virus or their methylation status,¹⁰⁴ and the frequency of such variants differs between ethnicities.^{104–108} Another protein is the protease TMPRSS2, which cleaves the spike protein, allowing fusion and cell entry, and there are variants of TMPRSS2 that differ between ethnicities.^{109,110} Virus properties affected by the D614G mutation may further have downstream effects on virus entry because of ethnically distinct differences (e.g., in the frequency of the alpha antitrypsin protease inhibitor).¹¹¹ Recently, a third category of genes has been found to differ between ethnicities.¹¹² The odorant metabolizing enzyme UGT2A1 which is expressed primarily in sustentacular cells and olfactory cilia¹¹³ was associated with COVID-related anosmia in a trans-ethnic analysis.¹¹² Such studies support the idea that host protein variants may contribute to the extent of anosmia and could explain, at least in part, ethnic differences in anosmia prevalence in COVID-19 (reviewed in Butowt et al., 2020).⁸

When using the same meta-analytic methods to compare cohorts primarily infected with the same virus strain (thus controlling for the virus type at a coarse level), the prevalence of olfactory dysfunction in the South Asian population was significantly lower than the prevalence reported for Caucasians ($p = 0.0054$, Figure 4B, D). The weighted random prevalence

of anosmia among Caucasian COVID patients was 49.02% (95% CI = 42.25–55.84%, $N = 77$) and 31.79% (95% CI = 23.26–41.76%, $N = 25$) among South Asian COVID patients. This suggests a relatively small, but measurable contribution of the host (ethnic difference in frequency of ACE2 variants or proteins such as UGT2A1).

Why Does SARS-CoV-2 Cause Much More Anosmia than SARS-CoV-1? The mechanism by which SARS-CoV-2 causes anosmia is beginning to come into view. Several different scenarios were initially considered: nasal obstruction (possibly due to inflammation), olfactory receptor neuron damage, olfactory support cell damage, and damage to central olfactory pathways.^{3,4,10,114,115} An emerging consensus favors a crucial role of the sustentacular support cells in the olfactory epithelium as the primary mechanism of COVID-induced anosmia.^{5,8,114,116–120} Since the SARS-CoV-2 entry protein (ACE2) is not or only minimally expressed in olfactory receptor neurons, the virus rarely infects the neurons,¹²¹ but rather enters the olfactory epithelium through sustentacular support cells and secretory cells in Bowman glands and primarily damages these cell types (Figure 5A). When

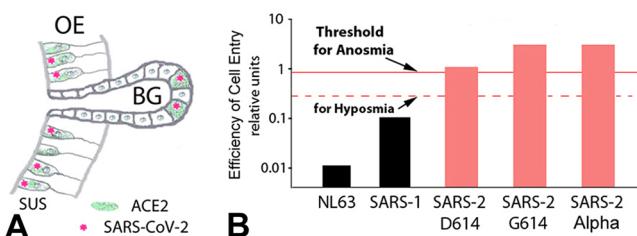


Figure 5. Illustration of the cell types infected in the olfactory epithelium (OE) (A) and the concept that the cell entry efficiency of the coronavirus determines the extent of damage that causes hyposmia or anosmia (B). This would explain the varying prevalence of olfactory dysfunction in patients with COVID-19 between populations and dominance of human coronaviruses NL63, SARS-1, SARS-2, and its strains D614, G614, or the G614 alpha variant. Efficiency of cell entry includes differences in binding affinities, fusion efficiency via the novel furin cleavage site, and neuropilin-1 binding as a cohost. For details, see Butowt et al., 2020.⁸ ACE2, angiotensin converting enzyme 2; BG, Bowman gland; SUS, sustentacular cell.

sustentacular cells become infected, they rapidly die, which appears to cause retraction of the adjacent neurons' ciliary processes,^{116,122} and may down-regulate expression of odorant receptors necessary for olfactory transduction.¹²³ In addition, infection and damage of Bowman gland cells may alter the composition of the mucus that is required for efficient access of odorants to the odorant receptors on the neuron's cilia.^{119,120} This prevents or alters binding of odorants to the olfactory neurons and thereby impacts olfactory transduction. After the degeneration of the sustentacular cells, stem cells in the olfactory epithelium divide (within a few days after death of sustentacular cells)¹¹⁴ and rapidly regenerate the lost sustentacular support cells,¹¹⁴ allowing the olfactory epithelium to be repaired. The neurons then recover, restore their cilia, resume odorant receptor expression, and the sense of smell returns in most cases within 1–2 weeks.¹ One study has proposed that the SARS-CoV-2 virus resembles olfactory receptors, and that IgA produced against the virus may thereby compromise olfaction as "collateral damage".¹²⁴ However, the early timing of loss of smell vs the delayed production of IgA makes this model an unlikely mechanism, although antigen exposure via

the olfactory epithelium may indeed boost the immune response and could lead to a more successful viral clearance and milder COVID-19 at the cost of a (temporary) loss of smell.¹²⁰

While the SARS-CoV-2 virus readily induces anosmia, the SARS-CoV-1 virus does not, even though the two virus types bind to the same entry receptor, ACE2.⁸ So what is different? The SARS-CoV-2 virus has a significantly higher cell entry efficiency than the SARS-CoV-1 virus (Figure 5B). In addition, the SARS-CoV-2 virus has a furin cleavage site that SARS-CoV-1 does not have.¹²⁵ By enhancing fusion efficiency, this site is thought to make SARS-CoV-2 more pathogenic.¹²⁶ The furin cleavage site may increase tropism (widening of the range of susceptible host cells) and cause higher pathogenicity.^{109,127} The higher binding affinity and the new furin cleavage site may both be responsible for the higher infectivity and higher anosmia prevalence of SARS-CoV-2 compared with SARS-CoV-1. Cleavage of the S1 spike protein generates a neuropilin-1 binding site, and therefore, neuropilin-1 can act as a host factor for SARS-CoV-2.^{128,129} Neuropilin is enriched in the olfactory epithelium,¹²⁸ indicating that the increased anosmia may involve binding to neuropilin-1.

Role of the D614G Mutation for Anosmia—Presumed Molecular Mechanisms. A key finding to understand how SARS-CoV-2 causes anosmia is that the mutated G614 virus is much more detrimental to olfaction than the original D614 virus. Why is the D614G mutation so much more effective in targeting the olfactory epithelium? What are the molecular mechanisms of this mutation? A number of studies have explored consequences of the D614G mutation, both for the clinical phenotype and for the pathophysiology at the molecular level. Clinically, despite the higher viral load, studies failed to detect an effect of the D614G mutation on the severity of COVID-19, hospitalization rate, or mortality.^{14,130,131}

Regarding the molecular mechanism of how the D614G mutation increases infectivity, transmission and possible disease severity, four different mechanisms are currently discussed, as recently reviewed.^{132–134} These are (1) modulation of the spike protein (by adding an elastase cleavage site or making furin cleavage more efficient);^{135,136} (2) promoting an open conformation of the receptor binding domain that favors ACE2 interaction;^{81,137} (3) increasing spike density and, therefore, facilitating cell entry;⁸¹ and (4) enhancing the stability of the spike protein (stronger retention of S1, less shedding of S1).^{13,138} All four mechanisms may contribute to enhanced cell entry and infectivity of the G614 virus.

But does this explain why the G614 virus is so much more effective at attacking the olfactory system? The G614 virus infects the upper respiratory tract, including nasal epithelium, more than lung epithelial cells, resulting in higher viral loads in the olfactory epithelium than in the lower respiratory tract.^{117,139} The enhanced cleavage at the furin cleavage site is of potential interest because the support cells and Bowman gland cells in the olfactory epithelium coexpress not only ACE2 and TMPRSS2 but also furin.¹⁴⁰ Accordingly, the cells in the olfactory epithelium may be more efficiently infected with the G614 virus, and this may explain the higher viral load in the nasal epithelium than in the lungs.¹¹¹ If the replicating virions are already cleaved by intracellular furin, as they exit the host cell, then they are ready to fuse with the next host cell, resulting in a more fulminant spread.¹⁴¹

On the other hand, the G614 mutation exposes the virus spike protein more than the D614 virus does, making the G614 virus more immunogenic, and possibly eliciting a stronger host immune response.^{11,120,137,142} Such differences in immunogenicity may indicate that the G614 virus, when it infects the nasal epithelium with a higher viral load, may trigger a more robust host immune defense. Although the virus may move to a new host too quickly to be affected by a neutralizing antibody response,¹³⁴ it cannot be ruled out a scenario where the virus in the olfactory epithelium (often preceding any other symptoms) elicits an immune host response that gives the host enough time to accelerate virus clearance and to prevent a subsequent more deadly infection of the lungs.¹²⁰ This may explain why SARS-CoV-2 infection leading to anosmia is associated with an overall milder COVID-19 disease, possibly because the nasal cavity-elicited immune defense leads to faster virus clearance¹⁴³ and thus reduces severe COVID-19 lung disease and death after G614 virus infection.^{144,145} Such a scenario may explain the puzzling finding that the G614 virus, despite being more infectious and leading to higher nasopharyngeal viral loads than the D614 virus, does not—overall—cause more severe and deadly COVID-19.

The presumed mechanisms of increasing efficiencies of binding and cell entry are summarized in Figure 5A and B. Regardless of the precise mechanism, our analysis shows an increased prevalence of olfactory dysfunction in cohorts infected with the G614 virus. Apparently, the G614 virus is more efficient than the D614 virus in entering and damaging the olfactory epithelium and impairing olfactory function. Importantly, our review identifies a “missing link” by revealing increased prevalence of chemosensory dysfunction as a novel, previously unrecognized phenotype of the now dominant G614 virus.

How Will Current and Future Novel Virus Variants Affect Olfactory Function? Since the spread of the G614 virus throughout most of the world, several new SARS-CoV-2 virus variants have emerged—Alpha (B.1.1.7), Beta (B.1.351), Gamma (B1.1.28), and Delta (B.1.617.2). All of these new variants also harbor the D614G mutation⁴ and, therefore, can be expected to cause similarly increased olfactory dysfunction as the G614 virus. These variants have additional spike protein mutations, besides the G614 mutation, resulting in already proven or suspected differences in their receptor binding properties, transmissibility, viral loads, and, in some cases, increased mortality.¹⁴⁶ For example, the virus with the N501Y mutation has similar binding to ACE2, while the K417N and the E484K mutants may have slightly increased binding to ACE2.¹⁴⁷ Multiple mutations can have interdependent and complex effects on binding and subsequent steps such as membrane fusion and host cell entry. For example, the Alpha variant that often also has N501Y, N439K, and Y453F mutations appears to require a deletion (Δ H69/V70) in the spike protein to maintain optimal cleavage and infectivity.¹⁴⁸ How these mutations generally affect infectivity *in vivo*, and specifically for cells in the olfactory epithelium, is not yet known.

The prevalence of olfactory dysfunction has been reported so far in only one of these variants (Alpha); it did not cause a significant change in anosmia prevalence.¹⁴⁹ An additional commentary that was based on apparently less reliable data reported an anosmia prevalence for the Alpha variant that differed by less than 4% from that of the G614 virus.^{150,151} Future studies will be needed for reliable and conclusive data

on the phenotypes of these new variants in terms of ACE2 binding affinity, membrane fusion, spike protein shedding, efficiency of host cell entry, viral load in different tissues, transmissibility, infectivity, mortality, and chemosensory dysfunction. Such phenotypes cannot be reliably predicted based on atomic modeling of the receptor binding domain of the spike protein because of the assumptions and restrictions of the modeling parameters.¹⁵² The cell entry properties of the new virus variants likely will continue to cause chemosensory dysfunction, and otorhinolaryngologists should expect to see such symptoms in COVID-19 patients when the new variants take over, as long as they maintain the D614G mutation, and assuming that the additional mutations do not neutralize their apparent effect on olfaction.

CONCLUSION

Our analysis shows an increased prevalence of olfactory dysfunction in COVID-19 patients infected with the D614G SARS-CoV-2 virus when compared with patients infected with the original D614 virus. Apparently, the G614 virus is more efficient than the D614 virus in entering and damaging the olfactory epithelium and impairing olfactory function. This finding has important implications for understanding the mechanism how SARS-CoV-2 compromises the olfactory system, and how future virus variants may similarly affect chemosensory function. Our analysis identifies enhanced prevalence of chemosensory dysfunction as a novel, previously unrecognized phenotype of the now dominant virus with the D614G mutation.

METHODS

For mapping and quantification of the contributions of D614 and G614 viruses in countries, we relied on the tracking Web site (<https://cov.lanl.gov/apps/covid-19/map/>)¹⁴ and for the regional geographical distribution within and outside of India we consulted multiple sources.^{14,16–22,24–29,31–36,100,153} Our study followed the PRISMA guidelines for systematic reviews and meta-analyses.⁴¹ We searched the COVID-19 portfolio of the National Institutes of Health (<https://icite.od.nih.gov/covid19/search/>) with the keywords, “anosmia” or “smell” and “India”, “Bangladesh”, “Pakistan” on and before July 22, 2021, resulting in 598 records. In addition, PubMed was searched (“anosmia”, “COVID”, “India”, “Bangladesh”, or “Pakistan”), resulting in 104 records. After removal of duplicates, 351 full-length texts were screened for the inclusion criteria: South Asian ethnicity (Indian, Pakistani, or Bangladeshi); a confirmed COVID-19 diagnosis; majority of subjects adults or teenagers; information about the weeks or months of the study periods; subjective (not objective) testing for olfactory dysfunction; we accepted no case reports, and included only reports of primary data, but no reviews. Studies were excluded when they lacked subjective testing,¹⁵⁴ when they considered only severe COVID cases,¹⁵⁵ reported on mostly East Asian rather than South Asian ethnicity,¹⁵⁶ or lacked necessary information about study periods, despite repeated requests.¹⁵⁷ The studies that met our inclusion criteria typically were cross-sectional, retrospective, observational studies that could be affected by recollection bias. However, such bias would be expected to be of a similar magnitude in studies examining patients infected with the D614 virus and studies examining patients infected with the G614 virus. We found 40 studies reporting on 43 cohorts that met our inclusion criteria. We compiled olfactory dysfunction regardless whether taste was also affected or not. The included studies are listed chronologically and sorted by D614 or G614 dominance in Table 1. A pooled analysis was performed for prevalence, and significance and confidence intervals were calculated in the software R, version 3.6.1 (R Foundation for Statistical Computing, Vienna, Austria). To calculate estimates of pooled prevalence and 95%

confidence intervals, we used the R-meta package, version 4.9–5, and the metaprop function. We used random effects models with the inverse variance method for pooling and the logit transformation for proportions.¹⁵⁸ For ease of interpretation, we back transformed and rescaled proportions to events per 100 observations. Subgroup analyses were conducted for binary group variables (D614 vs G614 and Caucasian vs South Asian) using the byvar statement of the metaprop function. All other subgroup tests used continuous variables and the metareg function, including the multivariable meta-regression. The subgroup age was a created variable that used the center of the sample, either the mean or the median, to mark the center of the age distribution, with the majority of studies reporting mean age (75.4%, $N = 32/43$). Analysis of the heterogeneity across studies was done using the Maximum-likelihood estimator, Higgins' I^2 , and Cochran's Q method.^{158,159} Publication bias was assessed by visual inspection of funnel plots¹⁶⁰ (Supporting Figures 2 and 3). In all cases, significance was defined at $\alpha = 0.05$.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acschemneuro.1c00542>.

Bubble plot showing details of the subgroup comparison with a gender effect on anosmia prevalence ([PDF](#))

Funnel plot for prevalence of anosmia in D614 and G614 cohorts of South Asians with COVID ([PDF](#))

Funnel plot for prevalence of anosmia in G614 cohorts of Caucasians with COVID ([PDF](#))

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All authors contributed to the conception and design of this project and the writing of the manuscript.

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■ REFERENCES

- (1) von Bartheld, C. S.; Hagen, M. M.; Butowt, R. Prevalence of Chemosensory Dysfunction in COVID-19 Patients: A Systematic Review and Meta-analysis Reveals Significant Ethnic Differences. *ACS Chem. Neurosci.* **2020**, *11* (19), 2944–2961.
- (2) Kim, J. W.; Han, S. C.; Jo, H. D.; Cho, S. W.; Kim, J. Y. Regional and Chronological Variation of Chemosensory Dysfunction in COVID-19: a Meta-Analysis. *J. Korean Med. Sci.* **2021**, *36* (4), e40.
- (3) Mutiawati, E.; Fahriani, M.; Mamada, S. S.; Fajar, J. K.; Frediansyah, A.; Maliga, H. A.; Ilmawan, M.; Emran, T. B.; Ophinni, Y.; Ichsan, I.; Musadir, N.; Rabaan, A. A.; Dhama, K.; Syahrul, S.; Nainu, F.; Harapan, H. Anosmia and dysgeusia in SARS-CoV-2 infection: incidence and effects on COVID-19 severity and mortality, and the possible pathobiology mechanisms - a systematic review and meta-analysis. *F1000Research* **2021**, *10*, 40.
- (4) Zeng, M.; Wang, D. Y.; Mullol, J.; Liu, Z. Chemosensory Dysfunction in Patients with COVID-19: What Do We Learn from the Global Outbreak? *Curr. Allergy Asthma Rep.* **2021**, *21* (2), 6.
- (5) Kumar, A. A.; Lee, S. W. Y.; Lock, C.; Keong, N. C. Geographical Variations in Host Predisposition to COVID-19 Related Anosmia, Ageusia, and Neurological Syndromes. *Front. Med.* **2021**, *8*, 661359.
- (6) Thakur, K.; Sagayaraj, A.; Prasad, K. C.; Gupta, A. Olfactory Dysfunction in COVID-19 Patients: Findings from a Tertiary Rural Centre. *Indian J. Otolaryngol Head Neck Surg.* **2021**, *1*–7.
- (7) Boscutti, A.; Delvecchio, G.; Pignoni, A.; Cereda, G.; Ciappolino, V.; Bellani, M.; Fusar-Poli, P.; Brambilla, P. Olfactory and gustatory dysfunctions in SARS-CoV-2 infection: A systematic review. *Brain Behav Immun Health* **2021**, *15*, No. 100268.
- (8) Butowt, R.; Bilinska, K.; von Bartheld, C. S. Chemosensory Dysfunction in COVID-19: Integration of Genetic and Epidemiological Data Points to D614G Spike Protein Variant as a Contributing Factor. *ACS Chem. Neurosci.* **2020**, *11* (20), 3180–3184.
- (9) Lechien, J. R.; Chiesa-Estomba, C. M.; De Santi, D. R.; Horoi, M.; Le Bon, S. D.; Rodriguez, A.; Dequanter, D.; Blebic, S.; El Afia, F.; Distinguin, L.; Chekkouri-Idrissi, Y.; Hans, S.; Delgado, I. L.; Calvo-Henriquez, C.; Lavigne, P.; Falanga, C.; Barillari, M. R.; Cammaroto, G.; Khalife, M.; Leich, P.; Souchay, C.; Rossi, C.; Journe, F.; Hsieh, J.; Edjlali, M.; Carlier, R.; Ris, L.; Lovato, A.; De Filippis, C.; Coppee, F.; Fakhry, N.; Ayad, T.; Saussez, S. Olfactory and gustatory dysfunctions as a clinical presentation of mild-to-moderate forms of the coronavirus disease (COVID-19): a multicenter European study. *Eur. Arch Otorhinolaryngol.* **2020**, *277* (8), 2251–2261.
- (10) Meng, X.; Deng, Y.; Dai, Z.; Meng, Z. COVID-19 and anosmia: A review based on up-to-date knowledge. *Am. J. Otolaryngol.* **2020**, *41* (5), 102581.
- (11) Yurkovetskiy, L.; Wang, X.; Pascal, K. E.; Tomkins-Tinch, C.; Nyatile, T. P.; Wang, Y.; Baum, A.; Diehl, W. E.; Dauphin, A.; Carbone, C.; Veinotte, K.; Egri, S. B.; Schaffner, S. F.; Lemieux, J. E.; Munro, J. B.; Rafique, A.; Barve, A.; Sabeti, P. C.; Kyratsous, C. A.; Dudkina, N. V.; Shen, K.; Luban, J. Structural and Functional Analysis of the D614G SARS-CoV-2 Spike Protein Variant. *Cell* **2020**, *183* (3), 739–751.
- (12) von Bartheld, C. S.; Mathew, D.; Butowt, R. New study on prevalence of anosmia in COVID-19 implicates the D614G virus mutation as a major contributing factor to chemosensory dysfunction. *Eur. Arch Otorhinolaryngol.* **2021**, *278* (9), 3593–3594.
- (13) Zhang, J.; Cai, Y.; Xiao, T.; Lu, J.; Peng, H.; Sterling, S. M.; Walsh, R. M., Jr; Rits-Volloch, S.; Zhu, H.; Woosley, A. N.; Yang, W.;

- Sliz, P.; Chen, B. Structural impact on SARS-CoV-2 spike protein by D614G substitution. *Science* **2021**, *372*, 525–530.
- (14) Korber, B.; Fischer, W. M.; Gnanakaran, S.; Yoon, H.; Theiler, J.; Abfaluterer, W.; Hengartner, N.; Giorgi, E. E.; Bhattacharya, T.; Foley, B.; Hastie, K. M.; Parker, M. D.; Partridge, D. G.; Evans, C. M.; Freeman, T. M.; de Silva, T. I. Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G Increases Infectivity of the COVID-19 Virus. *Cell* **2020**, *182* (4), 812–827.
- (15) Jacob, C. O. On the genetics and immunopathogenesis of COVID-19. *Clin. Immunol.* **2020**, *220*, 108591.
- (16) Bassa, B. V.; Uppu, R. M. The Frequency of G614 SARS-CoV-2 Variant in India. *Preprints*, 2020. DOI: [10.20944/preprints202008.0148.v1](https://doi.org/10.20944/preprints202008.0148.v1).
- (17) Kumar, B. K.; Venkatraja, B.; Prithvisagar, K. S.; Rai, P.; Rohit, A.; Hegde, M. N.; Karunasagar, I.; Karunasagar, I. Mutational analysis unveils the temporal and spatial distribution of G614 genotype of SARS-CoV-2 in different Indian states and its association with case fatality rate of COVID-19. *bioRxiv*, 2020. DOI: [10.1101/2020.07.27.222562](https://doi.org/10.1101/2020.07.27.222562).
- (18) Kumar, P.; Pandey, R.; Sharma, P.; Dhar, M. S.; A, V.; Uppili, B.; Vashisht, H.; Wadhwa, S.; Tyagi, N.; Fatihi, S.; Sharma, U.; Singh, P.; Lall, H.; Datta, M.; Gupta, P.; Saini, N.; Tewari, A.; Nandi, B.; Kumar, D.; Bag, S.; Gahlot, D.; Rathore, S.; Jatana, N.; Jaiswal, V.; Gogia, H.; Madan, P.; Singh, S.; Singh, P.; Dash, D.; Bala, M.; Kabra, S.; Singh, S.; Mukerji, M.; Thukral, L.; Faruq, M.; Agrawal, A.; Rakshit, P. Integrated genomic view of SARS-CoV-2 in India. *Wellcome Open Res.* **2020**, *5*, 184.
- (19) Maitra, A.; Raghav, S.; Dalal, A.; Ali, F.; Paynter, V. M.; Paul, D.; Biswas, N. K.; Ghosh, A.; Jani, K.; Chinnaswamy, S.; Pati, S.; Sahu, A.; Mitra, D.; Bhat, M. K.; Mayor, S.; Sarin, A.; The PANINDIA 1000 SARS-CoV-2 RNA Genome Sequencing Consortium; Sauche, Y. S.; Seshasayee, A. S. N.; Palakodeti, D.; Bashyam, M. D.; Parida, A.; Das, S. PAN-INDIA 1000 SARS-CoV-2 RNA genome sequencing reveals important insights into the outbreak. *bioRxiv*, 2020. DOI: [10.1101/2020.03.23.3718](https://doi.org/10.1101/2020.03.23.3718).
- (20) Raghav, S.; Ghosh, A.; Turuk, J.; Kumar, S.; Jha, A.; Madhulika, S.; Priyadarshini, M.; Biswas, V. K.; Shyamli, P. S.; Singh, B.; Singh, N.; Singh, D.; Datey, A.; Avula, K.; Smita, S.; Sabat, J.; Bhattacharya, D.; Kshatri, J. S.; Vasudevan, D.; Suryawanshi, A.; Dash, R.; Senapati, S.; Beuria, T. K.; Swain, R.; Chattopadhyay, S.; Syed, G. H.; Dixit, A.; Prasad, P.; et al. Odisha COVID-19 Study Group; ILS COVID-19 Team, Pati S, Parida A. Analysis of Indian SARS-CoV-2 Genomes Reveals Prevalence of D614G Mutation in Spike Protein Predicting an Increase in Interaction with TMPRSS2 and Virus Infectivity. *Front. Microbiol.* **2020**, *11*, 594928.
- (21) Samyuktha, V.; Naveen Kumar, V. Emergence of RBD and D614G Mutations in Spike Protein: An Insight from Indian SARS-CoV-2 Genome Analysis. *Preprints*, 2020, 2020060032, ver. 1. DOI: [10.20944/preprints202006.0032.v1](https://doi.org/10.20944/preprints202006.0032.v1).
- (22) Alai, S.; Gujar, N.; Joshi, M.; Gautam, M.; Gairola, S. Pan-India novel coronavirus SARS-CoV-2 genomics and global diversity analysis in spike protein. *Heliyon* **2021**, *7* (3), e06564.
- (23) See, A.; Ko, K. K. K.; Toh, S. T. Epidemiological analysis in support of hypothesis that D614G virus mutation is a major contributing factor to chemosensory dysfunction in COVID-19 patients. *Eur. Arch. Otorhinolaryngol.* **2021**, *278* (9), 3595–3596.
- (24) Srivastava, S.; Banu, S.; Singh, P.; Sowpati, D. T.; Mishra, R. K. SARS-CoV-2 genomics: An Indian perspective on sequencing viral variants. *J. Biosci.* **2021**, *46* (1), 22.
- (25) Yadav, P. D.; Nyayanit, D. A.; Majumdar, T.; Patil, S.; Kaur, H.; Gupta, N.; Shete, A. M.; Pandit, P.; Kumar, A.; Aggarwal, N.; Narayan, J.; Vijay, N.; Kalawat, U.; Sugunan, A. P.; Munivenkatappa, A.; Sharma, T.; Devi, S.; Majumdar, T.; Jaryal, S.; Bakshi, R.; Joshi, Y.; Sahay, R.; Shastri, J.; Singh, M.; Kumar, M.; Rawat, V.; Dutta, S.; Yadav, S.; Krishnasamy, K.; Raut, S.; Biswas, D.; Borkakoty, B.; Verma, S.; Rani, S.; Deval, H.; Patel, D.; Turuk, J.; Malhotra, B.; Fomda, B.; Nag, V.; Jain, A.; Bhargava, A.; Potdar, V.; Cherian, S.; Abraham, P.; Gopal, A.; Panda, S.; Bhargava, B. An Epidemiological Analysis of SARS-CoV-2 Genomic Sequences from Different Regions of India. *Viruses* **2021**, *13* (5), 925.
- (26) Islam, O. K.; Al-Emran, H. M.; Hasan, M. S.; Anwar, A.; Jahid, M. I. K.; Hossain, M. A. Emergence of European and North American mutant variants of SARS-CoV-2 in South-East Asia. *Transboundary Emerging Dis.* **2021**, *68* (2), 824–832.
- (27) Mondal, M.; Lawarde, A.; Somasundaram, K. Genomics of Indian SARS-CoV-2: Implications in genetic diversity, possible origin and spread of virus. *medRxiv*, 2020. DOI: [10.1101/2020.04.25.20079475](https://doi.org/10.1101/2020.04.25.20079475)
- (28) Pattabiraman, C.; Habib, F.; Rasheed, R.; Prasad, P.; Reddy, V.; Dinesh, P.; Damodar, T.; Hosallimath, K.; George, A. K.; Kiran Reddy, N. V.; John, B.; Pattanaik, A.; Kumar, N.; Mani, R. S.; Venkataswamy, M. M.; Shahul Hameed, S. K.; Desai, A.; Vasanthaapuram, R.; et al. Genomic epidemiology reveals multiple introductions and spread of SARS-CoV-2 in the Indian state of Karnataka. *PLoS One* **2020**, *15* (12), e0243412.
- (29) Sarkar, R.; Mitra, S.; Chandra, P.; Saha, P.; Banerjee, A.; Dutta, S.; Chawla-Sarkar, M. Comprehensive analysis of genomic diversity of SARS-CoV-2 in different geographic regions of India: an endeavour to classify Indian SARS-CoV-2 strains on the basis of co-existing mutations. *Arch. Virol.* **2021**, *166* (3), 801–812.
- (30) Gupta, A.; Sabarinathan, R.; Bala, P.; Donipadi, V.; Vashisht, D.; Katika, M. R.; Kandakatla, M.; Mitra, D.; Dalal, A.; Bashyam, M. D. A comprehensive profile of genomic variations in the SARS-CoV-2 isolates from the state of Telangana, India. *J. Gen. Virol.* **2021**, DOI: [10.1099/jgv.0.001562](https://doi.org/10.1099/jgv.0.001562).
- (31) Paul, D.; Jani, K.; Kumar, J.; Chauhan, R.; et al. Phylogenomic analysis of SARS-CoV-2 genomes from Western India reveals unique linked mutations. *bioRxiv*, 2020. DOI: [10.1101/2020.07.30.228460](https://doi.org/10.1101/2020.07.30.228460).
- (32) Hasan, M. M.; Das, R.; Rasheduzzaman, M.; Hussain, M. H.; Muzahid, N. H.; Salauddin, A.; Rumi, M. H.; Mahbubur Rashid, S. M.; Siddiki, A. Z.; Mannan, A. Global and local mutations in Bangladeshi SARS-CoV-2 genomes. *Virus Res.* **2021**, *297*, 198390.
- (33) Shishir, T. A.; Naser, I. B.; Faruque, S. M. In silico comparative genomics of SARS-CoV-2 to determine the source and diversity of the pathogen in Bangladesh. *PLoS One* **2021**, *16* (1), e0245584.
- (34) Ghanchi, N. K.; Masood, K. I.; Nasir, A.; Khan, W.; Abidi, S. H.; Shahid, S.; Mahmood, S. F.; Kanji, A.; Razzak, S.; Ansar, Z.; Islam, N.; Dharejo, M. B.; Hasan, Z.; Hasan, R. SARS-CoV-2 genome analysis of strains in Pakistan reveals GH, S and L clade strains at the start of the pandemic. *bioRxiv* **2020**, DOI: [10.1101/2020.08.04.234153](https://doi.org/10.1101/2020.08.04.234153).
- (35) Jeewandara, C.; Jayathilaka, D.; Ranasinghe, D.; Hsu, N. S.; Ariyaratne, D.; Jayadas, T. T.; Madushanka, D.; Lindsey, B. B.; Gomes, L.; Parker, M. D.; Wijewickrama, A.; Karunaratne, M.; Ogg, G. S.; de Silva, T. I.; Malavige, G. N. Genomic and epidemiological analysis of SARS-CoV-2 viruses in Sri Lanka. *medRxiv* **2021**, DOI: [10.1101/2021.05.05.21256384](https://doi.org/10.1101/2021.05.05.21256384).
- (36) Umair, M.; Ikram, A.; Salman, M.; Khurshid, A.; Alam, M.; Badar, N.; Suleman, R.; Tahir, F.; Sharif, S.; Montgomery, J.; Whitmer, S.; Klena, J. Whole-genome sequencing of SARS-CoV-2 reveals the detection of G614 variant in Pakistan. *PLoS One* **2021**, *16* (3), e0248371.
- (37) Sallam, M.; Ababneh, N. A.; Dababseh, D.; Bakri, F. G.; Mahafzah, A. Temporal increase in D614G mutation of SARS-CoV-2 in the Middle East and North Africa. *Heliyon* **2021**, *7* (1), e06035.
- (38) Ramasamy, K.; Saniasiaya, J.; Abdul Gani, N. Olfactory and Gustatory Dysfunctions as a Clinical Manifestation of Coronavirus Disease 2019 in a Malaysian Tertiary Center. *Ann. Otol., Rhinol., Laryngol.* **2021**, *130* (5), S13–S19.
- (39) Mat Yassim, A. S.; Asras, M. F. F.; Gazali, A. M.; Marcial-Coba, M. S.; Zainulabid, U. A.; Ahmad, H. F. COVID-19 Outbreak in Malaysia: Decoding D614G Mutation of SARS-CoV-2 Virus Isolated from an Asymptomatic Case in Pahang. *Mater. Today Proc.* **2021**, DOI: [10.1016/j.matpr.2021.02.387](https://doi.org/10.1016/j.matpr.2021.02.387).
- (40) Al-Mahruqi, S.; Al-Wahaibi, A.; Khan, A. L.; Al-Jardani, A.; Asaf, S.; Alkindi, H.; Al-Kharusi, S.; Al-Rawahi, A. N.; Al-Rawahi, A.; Al-Salmani, M.; Al-Shukri, I.; Al-Busaidi, A.; Al-Abri, S. S.; Al-Harrasi,

- A. Molecular epidemiology of COVID-19 in Oman: A molecular and surveillance study for the early transmission of COVID-19 in the country. *Int. J. Infect. Dis.* **2021**, *104*, 139–149.
- (41) Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D. G. PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* **2009**, *6* (7), e1000097.
- (42) Almazedi, S.; Al-Youha, S.; Jamal, M. H.; Al-Haddad, M.; Al-Muhaini, A.; Al-Ghimplas, F.; Al-Sabah, S. Characteristics, risk factors and outcomes among the first consecutive 1096 patients diagnosed with COVID-19 in Kuwait. *EClinicalMedicine* **2020**, *24*, 100448.
- (43) Tham, A. C.; Thein, T. L.; Lee, C. S.; Tan, G. S. E.; Manauis, C. M.; Siow, J. K.; Leo, Y. S.; Lim, M. Y. Olfactory taste disorder as a presenting symptom of COVID-19: a large single-center Singapore study. *Eur. Arch. Otorhinolaryngol.* **2021**, *278* (6), 1853–1862.
- (44) Soh, S. H. L.; See, A.; Teo, N. W. Y.; Tan, H. K.; Palaniappan, G.; Lim, M. L. A.; Kadir, H. B. A.; Toh, S. T. Prevalence of olfactory and taste dysfunction in COVID-19 patients: a community care facility study. *Eur. Arch. Otorhinolaryngol.* **2021**, *278* (9), 3375–3380.
- (45) Varghese, B.; Shajahan, S.; Anilkumar, H.; Haridasan, R. K.; Rahul, A.; Thazhathethadath, H.; Surendran Nair, A. T. Symptomatology and Epidemiologic Characteristics of COVID 19 Patients in Kerala. *J. Evol. Med. Dent. Sci.* **2020**, *9* (46), 3411–3417.
- (46) Ish, P.; Sen, M. K.; Gupta, N. In Reference to Anosmia and Ageusia: Common Findings in COVID-19 Patients. *Laryngoscope* **2020**, *130* (9), E502.
- (47) Sharma, A. K.; Ahmed, A.; Baig, V. N.; Dhakar, P.; Dalela, G.; Kacker, S.; Panwar, V. R.; Panwar, R. B.; Gupta, R. Characteristics and Outcomes of Hospitalized Young Adults with Mild Covid-19. *J. Assoc. Physicians India* **2020**, *68* (8), 62–65.
- (48) Herath, M. et al. Clinical and Epidemiological Characteristics and Outcome of Patients with Covid-19 in Sri Lanka: An Observational Study. *Research Square*, 2020. DOI: [10.21203/rs.3.rs-42505/v1](https://doi.org/10.21203/rs.3.rs-42505/v1).
- (49) Iftaf, S., Sr; Fatima, M.; Salman, S., Sr; Salam, J. U.; Abbas, S. Frequency of Neurological Presentations of Coronavirus Disease in Patients Presenting to a Tertiary Care Hospital During the 2019 Coronavirus Disease Pandemic. *Cureus* **2020**, *12* (8), e9846.
- (50) Lal, P.; Chamoli, P.; Tuli, I. P.; Jaitly, S.; Sneha, S. N.; Sharma, S.; Trehan, S. Olfactory and Gustatory Dysfunctions in Patients with Laboratory-Confirmed COVID-19 Infection: A Change in the Trend. *Indian J. Otolaryngol Head Neck Surg.* **2021**, 1–7.
- (51) Yadav, V.; Bhagat, S.; Sharma, D. K.; Sibia, R. P. S.; Pandav, R.; Sandhu, V. P. Olfactory and Gustatory Dysfunction in COVID-19 Patients: A Tertiary Care Institute Experience in India. *Indian J. Otolaryngol Head Neck Surg.* **2021**, 1–8.
- (52) Khurana, A.; Kaushal, G.; Gupta, R.; Verma, V.; Sharma, K.; Kohli, P. Prevalence and Clinical Correlates of COVID-19 Outbreak Among Health Care Workers in a Tertiary Level Hospital in Delhi. *Am. J. Infect. Dis.* **2021**, *17* (2), 107–119.
- (53) Lanjiwar, M.; Narayan SJ, A.; Garg, R.; Srivastava, P.; Vibha, D. Neurological manifestations in mild SARS-CoV2 infected patients at a tertiary health center in India: A prospective cohort study. SSRN, 2020. DOI: [10.2139/ssrn.3605278](https://doi.org/10.2139/ssrn.3605278)
- (54) Krishnasamy, N.; Natarajan, M.; Ramachandran, A.; Vivian Thangaraj, J. W.; Etherajan, T.; Rengarajan, J.; Shanmugasundaram, M.; Kandasamy, A.; Ramamoorthy, R.; Velusamy, A.; Obla Lakshmanamoorthy, N. B.; Kanagaraman, P.; Rahamathula, M. I.; Devadas, G.; Sathyananthan, B. P.; Rajaji, P.; Rajendran, K.; Panneerselvam, P.; Rajaram, M.; Panjacharam, M. Clinical Outcomes among Asymptomatic or Mildly Symptomatic COVID-19 Patients in an Isolation Facility in Chennai, India. *Am. J. Trop. Med. Hyg.* **2021**, *104* (1), 85–90.
- (55) Smitha, S. G.; Pillai, N.; Nayak, B.; Raveendran, J. A Study on Otorhinolaryngological Presentations in Covid 19 Patients in a Tertiary Health Care Center. *Indian J. Otolaryngol Head Neck Surg.* **2021**, DOI: [10.1007/s12070-021-02564-2](https://doi.org/10.1007/s12070-021-02564-2).
- (56) Makda, A.; Kumar, S.; Kumar, A.; Kumar, V.; Rizwan, A. The Frequency of Neurological Symptoms in COVID-19 Patients at a Tertiary Care Hospital in Pakistan. *Cureus* **2020**, *12* (9), No. e10360.
- (57) Al Harthi, S.; Al Osali, M.; Al Ismaili, R.; Al Lawati, S.; Kamble, B.; Al Shaibbi, M.; Al Kindi, N.; Al Qasabi, S.; Alhinai, M.; Al Harthi, H.; Al Ghafri, T. Clinical characteristics of confirmed cases of COVID-19 admitted at Al-Nahdha hospital, Oman: A cross-sectional descriptive study. *Cureus* **2021**, *13* (8), No. e17343.
- (58) Kuchhal, V.; Ahmad, S.; Chaurasia, P.; Rawat, P. ENT manifestations in COVID-19 positive patients. *Research Square* **2020**, DOI: [10.21203/rs.3.rs-90203/v1](https://doi.org/10.21203/rs.3.rs-90203/v1).
- (59) Mishra, P.; Gowda, V.; Dixit, S.; Kaushik, M. Prevalence of New Onset Anosmia in COVID-19 Patients: Is the Trend Different Between European and Indian Population? *Indian J. Otolaryngol Head Neck Surg.* **2020**, *72* (4), 484.
- (60) Bhatta, S.; Gandhi, S.; Saindani, S. J.; Ganesuni, D.; Ghanpur, A. D. Otorhinolaryngological manifestations of coronavirus disease 2019: a prospective review of 600 patients. *J. Laryngol. Otol.* **2021**, *135* (3), 206–211.
- (61) Panda, S.; Mohamed, A.; Sikka, K.; Kanodia, A.; Sakthivel, P.; Thakar, A.; Bhatnagar, S.; Mohan, A.; Meena, V. P.; Tiwari, P.; Sahoo, B.; Dar, L.; Vig, S.; Garg, R.; Kumar, C. Otolaryngologic Manifestation and Long-Term Outcome in Mild COVID-19: Experience from a Tertiary Care Centre in India. *Indian J. Otolaryngol Head Neck Surg.* **2021**, *73* (1), 72.
- (62) Chaurasia, P.; Kuchhal, V.; Ahmad, S.; Rawat, P. ENT manifestations in Covid-19 positive patients. *Int. J. Health Clin Res.* **2020**, *3* (10), 187–191.
- (63) Malik, F. T.; Ishraquzzaman, M.; Kalimuddin, M.; Choudhury, S.; Ahmed, N.; Badiuzzaman, M.; Ahmed, M. N.; Banik, D.; Huq, T. S.; Al Mamun, M. A. Clinical Presentation, Management and In-Hospital Outcome of Healthcare Personnel With COVID-19 Disease. *Cureus* **2020**, *12* (8), e10004.
- (64) Shah, N. N.; Hussain, R. T.; Mustafa, H.; Mushtaq, M.; Ali, M. Evaluation of Olfactory Acuity in Patients with Coronavirus Disease 2019 (COVID-19). *Indian J. Otolaryngol Head Neck Surg.* **2020**, 1–8.
- (65) Bhattacharjee, A. S.; Joshi, S. V.; Naik, S.; Sangle, S.; Abraham, N. M. Quantitative assessment of olfactory dysfunction accurately detects asymptomatic COVID-19 carriers. *EClinicalMedicine* **2020**, *28*, 100575.
- (66) Jain, A.; Kumar, L.; Kaur, J.; Baisla, T.; Goyal, P.; Pandey, A. K.; Das, A.; Parashar, L. Olfactory and taste dysfunction in coronavirus disease 2019 patients: its prevalence and outcomes. *J. Laryngol. Otol.* **2020**, *134*, 987–991.
- (67) Dev, N.; Sankar, J.; Gupta, N.; Meena, R. C.; Singh, C.; Gupta, D. K.; Sen, M. K. COVID-19 with and without anosmia or dysgeusia: A case-control study. *J. Med. Virol.* **2021**, *93* (4), 2499–2504.
- (68) Kumar, L.; Kahlon, N.; Jain, A.; Kaur, J.; Singh, M.; Pandey, A. K. Loss of smell and taste in COVID-19 infection in adolescents. *Int. J. Pediatr. Otorhinolaryngol.* **2021**, *142*, 110626.
- (69) Koul, D.; Begh, R. A.; Kalsotra, P. Olfactory and Gustatory Alterations in Covid-19 Patients: A Tertiary Care Covid-19 Centre Inpatient Experience. *Indian J. Otolaryngol Head Neck Surg.* **2021**, 1–5.
- (70) Sahoo, P. R.; Sahu, M.; Surapaneni, P. S.; Maiti, A.; Vankamamidi, R.; Panda, N.; Biswal, R. N. Evolution of olfactory and gustatory dysfunctions in COVID-19 patients in India. *Eur. Arch. Otorhinolaryngol.* **2021**, *278* (8), 2875–2881.
- (71) Rajkumar, I.; Anand, K. H.; Revathishree, K.; Shoba, K.; Srinivasan, K. Contemporary Analysis of Olfactory Dysfunction in Mild to Moderate Covid 19 Patients in A Tertiary Health Care Centre. *Indian J. Otolaryngol Head Neck Surg.* **2020**, 1–5.
- (72) Kumar, V.; Singla, S.; Gupta, N.; Bharati, S. J.; Garg, R.; Pandit, A.; Vig, S.; Mishra, S.; Bhatnagar, S. The incidence of anosmia in patients with laboratory-confirmed COVID 19 infection in India: An observational study. *J. Anaesthesiol Clin Pharmacol.* **2021**, *37* (1), 51–56.
- (73) Kandakure, V. T.; Valvi, H. R.; Khokle, P.; More, M. S.; Chouhan, R. Prevalence and Recovery from Newly Onset Anosmia and Ageusia in Covid 19 Patients at our Tertiary Care Centre. *Indian J. Otolaryngol Head Neck Surg.* **2021**, 1–8.

- (74) Hasan, M.; Tamanna, N. A.; Jamal, M. N.; Uddin, J. The prevalence of olfactory dysfunction and its associated factors in patients with COVID-19 infection. *medRxiv*, 2021. DOI: 10.1101/2021.01.27.21250153.
- (75) Hajare, P. S.; Harugop, A. S.; Goswami, L.; Padmavathy, O.; Aggarwal, U.; Reddy, Y. L. Prevalence of Olfactory and Gustatory Dysfunction in Coronavirus Disease (COVID-19): A Cross Sectional Study in our Tertiary Care Hospital. *Indian J. Otolaryngol Head Neck Surg.* 2021, 1–4.
- (76) Savtale, S.; Hippargekar, P.; Bhise, S.; Kothule, S. Prevalence of Otorhinolaryngological Symptoms in Covid 19 Patients. *Indian J. Otolaryngol Head Neck Surg.* 2021, 1–7.
- (77) Gupta, S.; Kumbhat, P.; Seervi, M. Olfactory and Gustatory Dysfunction in Covid-19: An Observational Study in a Tertiary Care Institute of Western Rajasthan. *Indian J. Otolaryngol Head Neck Surg.* 2021, 1–5.
- (78) Mangal, V.; Murari, T.; Vashisht, R.; Iqbal, S. M.; Meghana, K.; Gujrathi, S.; Ambade, V.; Tilak, T.; Aggarwal, V.; Manrai, M.; Verma, V.; Srinath, R.; Goel, N.; Yadav, N. K.; Menon, A. Olfactory Dysfunction Among Asymptomatic Patients with SARS CoV2 Infection: A Case-Control Study. *Indian J. Otolaryngol Head Neck Surg.* 2021, 73, 212–217.
- (79) Karthikeyan, P.; Sivanand, N.; Vijayan, N.; Latheef, M. N. A Clinical Study of Smell Disorders in COVID-19 Patients in a Tertiary Care Hospital in Pondicherry: A Cross Sectional Study. *Indian J. Otolaryngol Head Neck Surg.* 2021, 1–6.
- (80) Silu, M.; Mathur, N. P.; Kumari, R.; Chaudhary, P. Correlation Between Anosmia and Severity Along with Requirement of Tocilizumab in COVID-19 Patients. *Indian J. Otolaryngol Head Neck Surg.* 2021, 73 (3), 378–382.
- (81) Zhang, L.; Jackson, C. B.; Mou, H.; Ojha, A.; Peng, H.; Quinlan, B. D.; Rangarajan, E. S.; Pan, A.; Vanderheiden, A.; Suthar, M. S.; Li, W.; Izard, T.; Rader, C.; Farzan, M.; Choe, H. SARS-CoV-2 spike-protein D614G mutation increases virion spike density and infectivity. *Nat. Commun.* 2020, 11 (1), 6013.
- (82) Agyeman, A. A.; Chin, K. L.; Landersdorfer, C. B.; Liew, D.; Ofori-Asenso, R. Smell and Taste Dysfunction in Patients with COVID-19: A Systematic Review and Meta-analysis. *Mayo Clin. Proc.* 2020, 95 (8), 1621–1631.
- (83) Fuccillo, E.; Saibene, A. M.; Canevini, M. P.; Felisati, G. Olfactory disorders in coronavirus disease 2019 patients: a systematic literature review. *J. Laryngol. Otol.* 2020, 134, 754–763.
- (84) Printza, A.; Constantinidis, J. The role of self-reported smell and taste disorders in suspected COVID-19. *Eur. Arch. Otorhinolaryngol.* 2020, 277 (9), 2625–2630.
- (85) Purja, S.; Shin, H.; Lee, J. Y.; Kim, E. Is loss of smell an early predictor of COVID-19 severity: a systematic review and meta-analysis. *Arch. Pharmacal Res.* 2021, 44 (7), 725–740.
- (86) Aziz, M.; Goyal, H.; Haghbin, H.; Lee-Smith, W. M.; Gajendran, M.; Perisetti, A. The Association of “Loss of Smell” to COVID-19: A Systematic Review and Meta-Analysis. *Am. J. Med. Sci.* 2021, 361 (2), 216–225.
- (87) Tong, J. Y.; Wong, A.; Zhu, D.; Fastenberg, J. H.; Tham, T. The Prevalence of Olfactory and Gustatory Dysfunction in COVID-19 Patients: A Systematic Review and Meta-analysis. *Otolaryngol.–Head Neck Surg.* 2020, 163 (1), 3–11.
- (88) Wong, D. K. C.; Gendeh, H. S.; Thong, H. K.; Lum, S. G.; Gendeh, B. S.; Saim, A.; Salina, H. A review of smell and taste dysfunction in COVID-19 patients. *Med. J. Malaysia* 2020, 75 (5), 574–581.
- (89) Lee, Y.; Min, P.; Lee, S.; Kim, S. W. Prevalence and Duration of Acute Loss of Smell or Taste in COVID-19 Patients. *J. Korean Med. Sci.* 2020, 35 (18), e174.
- (90) Wee, L. E.; Chan, Y. F. Z.; Teo, N. W. Y.; Cherng, B. P. Z.; Thien, S. Y.; Wong, H. M.; Wijaya, L.; Toh, S. T.; Tan, T. T. The role of self-reported olfactory and gustatory dysfunction as a screening criterion for suspected COVID-19. *Eur. Arch. Otorhinolaryngol.* 2020, 277 (8), 2389–2390.
- (91) Qiu, C.; Cui, C.; Hautefort, C.; Haehner, A.; Zhao, J.; Yao, Q.; Zeng, H.; Nisenbaum, E. J.; Liu, L.; Zhao, Y.; Zhang, D.; Levine, C. G.; Cejas, L.; Dai, Q.; Zeng, M.; Herman, P.; Jourdain, C.; de With, K.; Draf, J.; Chen, B.; Jayaweera, D. T.; Denneny, J. C., 3rd; Casiano, R.; Yu, H.; Eshraghi, A. A.; Hummel, T.; Liu, X.; Shu, Y.; Lu, H. Olfactory and Gustatory Dysfunction as an Early Identifier of COVID-19 in Adults and Children: An International Multicenter Study. *Otolaryngol.–Head Neck Surg.* 2020, 163 (4), 714–721.
- (92) Song, J.; Deng, Y. K.; Wang, H.; Wang, Z. C.; Liao, B.; Ma, J.; He, C.; Pan, L.; Liu, Y.; Alobid, I.; Wang, D. Y.; Zeng, M.; Mullol, J.; Liu, Z. Self-reported Taste and Smell Disorders in Patients with COVID-19: Distinct Features in China. *Curr. Med. Sci.* 2021, 41 (1), 14–23.
- (93) Hannum, M. E.; Ramirez, V. A.; Lipson, S. J.; Herriman, R. D.; Toskala, A. K.; Lin, C.; Joseph, P. V.; Reed, D. R. Objective Sensory Testing Methods Reveal a Higher Prevalence of Olfactory Loss in COVID-19-Positive Patients Compared to Subjective Methods: A Systematic Review and Meta-Analysis. *Chem. Senses* 2020, 45 (9), 865–874.
- (94) Pang, K. W.; Chee, J.; Subramaniam, S.; Ng, C. L. Frequency and Clinical Utility of Olfactory Dysfunction in COVID-19: a Systematic Review and Meta-analysis. *Curr. Allergy Asthma Rep.* 2020, 20 (12), 76.
- (95) Aanand, P.; Angral, S.; Varshney, S.; Raj, R. Incidence of Anosmia among Covid 19 patients in India. *Indian J. Otolaryngol Head Neck Surg.* 2021.
- (96) Eyre, D. W.; Lumley, S. F.; O'Donnell, D.; Stoesser, N. E.; Matthews, P. C.; Howarth, A.; Hatch, S. B.; Marsden, B. D.; Cox, S.; James, T.; Cornall, R. J.; Stuart, D. I.; Scream, G.; Ebner, D.; Crook, D. W.; Conlon, C. P.; Jeffery, K.; Walker, T. M.; Peto, T. E. A. Stringent thresholds in SARS-CoV-2 IgG assays lead to under-detection of mild infections. *BMC Infect. Dis.* 2021, 21 (1), 187.
- (97) Chung, T. W.; Sridhar, S.; Zhang, A. J.; Chan, K. H.; Li, H. L.; Wong, F. K.; Ng, M. Y.; Tsang, R. K.; Lee, A. C.; Fan, Z.; Ho, R. S.; Luk, S. Y.; Kan, W. K.; Lam, S. H.; Wu, A. K.; Leung, S. M.; Chan, W. M.; Ng, P. Y.; To, K. K.; Cheng, V. C.; Lung, K. C.; Hung, I. F.; Yuen, K. Y. Olfactory Dysfunction in Coronavirus Disease 2019 Patients: Observational Cohort Study and Systematic Review. *Open Forum Infect Dis.* 2020, 7 (6), No. ofaa199.
- (98) Cho, R. H. W.; To, Z. W. H.; Yeung, Z. W. C.; Tso, E. Y. K.; Fung, K. S. C.; Chau, S. K. Y.; Leung, E. Y. L.; Hui, T. S. C.; Tsang, S. W. C.; Kung, K. N.; Chow, E. Y. D.; Abdullah, V.; van Hasselt, A.; Tong, M. C. F.; Ku, P. K. M. COVID-19 Viral Load in the Severity of and Recovery from Olfactory and Gustatory Dysfunction. *Laryngoscope* 2020, 130 (11), 2680–2685.
- (99) Leung, W. L. H.; Yu, E. L. M.; Wong, S. C.; Leung, M.; Lee, L. L. Y.; Chung, K. L.; Cheng, V. C. C. Findings from the first public COVID-19 temporary test centre in Hong Kong. *Hong Kong Med. J.* 2021, 27 (2), 99–105.
- (100) Mak, G. C. K.; Lau, A. W. L.; Chan, A. M. Y.; Chan, D. Y. W.; Tsang, D. N. C. The D614G substitution in the S gene and clinical information for patients with COVID-19 detected in Hong Kong. *J. Clin. Virol.* 2020, 130, 104550.
- (101) Lee, S.-H.; Yeoh, Z. X.; Sachlin, I. S.; Gazali, N.; Soelar, S. A.; Foo, C. Y.; Low, L. L.; Alwai, S. B. S.; Kamaldien, T. M. I. T.; Shanmuganathan, J.; et al.; Malaysian COVID-19 Anosmia Research Group. Self-reported symptom study of COVID-19 chemosensory dysfunction in Malaysia. *Research Square*, 2021. DOI: 10.21203/rs.3.rs-696505/v1
- (102) Gudbjartsson, D. F.; Helgason, A.; Jonsson, H.; Magnusson, O. T.; Melsted, P.; Norddahl, G. L.; Saemundsdottir, J.; Sigurdsson, A.; Sulem, P.; Agustsdottir, A. B.; Eiriksdottir, B.; Fridriksdottir, R.; Gardarsdottir, E. E.; Georgsson, G.; Gretarsdottir, O. S.; Gudmundsson, K. R.; Gunnarsdottir, T. R.; Gylfason, A.; Holm, H.; Jensson, B. O.; Jonasdottir, A.; Jonsson, F.; Josefsdottir, K. S.; Kristjansson, T.; Magnusdottir, D. N.; le Roux, L.; Sigmundsdottir, G.; Sveinbjornsson, G.; Sveinsdottir, K. E.; Sveinsdottir, M.; Thorarensen, E. A.; Thorbjornsson, B.; Löve, A.; Masson, G.; Jonsdottir, I.; Möller, A. D.; Gudnason, T.; Kristinsson, K. G.;

- Thorsteinsdottir, U.; Stefansson, K. Spread of SARS-CoV-2 in the Icelandic Population. *N. Engl. J. Med.* **2020**, *382* (24), 2302–2315.
- (103) Borobia, A. M.; Carcas, A. J.; Arnalich, F.; Alvarez-Sala, R.; Monserrat-Villatoro, J.; Quintana, M.; Figueira, J. C.; Torres Santos-Olmo, R. M.; García-Rodríguez, J.; Martín-Vega, A.; Buño, A.; Ramírez, E.; Martínez-Alés, G.; García-Arenzana, N.; Núñez, M. C.; Martí-de-Gracia, M.; Moreno Ramos, F.; Reinoso-Barbero, F.; Martin-Quiros, A.; Rivera Núñez, A.; Mingorance, J.; Carpio Segura, C. J.; Prieto Arribas, D.; Rey Cuevas, E.; Prados Sánchez, C.; Rios, J. J.; Hernán, M. A.; Frías, J.; Arribas, J. R. A Cohort of Patients with COVID-19 in a Major Teaching Hospital in Europe. *J. Clin. Med.* **2020**, *9* (6), 1733.
- (104) Cardenas, A.; Rifas-Shiman, S. L.; Sordillo, J. E.; DeMeo, D. L.; Baccarelli, A. A.; Hivert, M. F.; Gold, D. R.; Oken, E. DNA methylation architecture of the ACE2 gene in nasal cells of children. *Sci. Rep.* **2021**, *11* (1), 7107.
- (105) Asselta, R.; Paraboschi, E. M.; Mantovani, A.; Duga, S. ACE2 and TMPRSS2 variants and expression as candidates to sex and country differences in COVID-19 severity in Italy. *Aging* **2020**, *12* (11), 10087–10098.
- (106) Cao, Y.; Li, L.; Feng, Z.; Wan, S.; Huang, P.; Sun, X.; Wen, F.; Huang, X.; Ning, G.; Wang, W. Comparative genetic analysis of the novel coronavirus (2019-nCoV/SARS-CoV-2) receptor ACE2 in different populations. *Cell Discovery* **2020**, *6*, 11.
- (107) Strafella, C.; Caputo, V.; Termine, A.; Barati, S.; Gambardella, S.; Borgiani, P.; Caltagirone, C.; Novelli, G.; Giardina, E.; Cascella, R. Analysis of ACE2 Genetic Variability among Populations Highlights a Possible Link with COVID-19-Related Neurological Complications. *Genes* **2020**, *11* (7), 741.
- (108) Williams, F. M. K.; Freidin, M. B.; Mangino, M.; Couvreur, S.; Visconti, A.; Bowyer, R. C. E.; Le Roy, C. I.; Falchi, M.; Mompeó, O.; Sudre, C.; Davies, R.; Hammond, C.; Menni, C.; Steves, C. J.; Spector, T. D. Self-Reported Symptoms of COVID-19, Including Symptoms Most Predictive of SARS-CoV-2 Infection, Are Heritable. *Twin Res. Hum. Genet.* **2020**, *23* (6), 316–321.
- (109) Bhattacharyya, C.; Das, C.; Ghosh, A.; Singh, A. K.; Mukherjee, S.; Majumder, P. P.; Basu, A.; Biswas, N. K. Global Spread of SARS-CoV-2 Subtype with Spike Protein Mutation D614G is Shaped by Human Genomic Variations that Regulate Expression of TMPRSS2 and MX1 Genes. *bioRxiv*, 2020. DOI: [10.1101/2020.05.04.075911](https://doi.org/10.1101/2020.05.04.075911).
- (110) dos Santos, N. P. C.; Khayat, A. S.; Rodrigues, J. C. G.; Pinto, P.; de Araújo, G. S.; Pastana, L. F.; Medeiros, J. A. G.; Fernandes, M. R.; Ribeiro-dos-Santos, A.; Khayat, B. C. M.; Cordeiro Moreira, F. C.; Ribeiro-dos-Santos, A. M.; de Assumpção, P. B.; Ribeiro-dos-Santos, A.; de Assumpção, P. P.; Santos, S. TMPRSS2 variants and their susceptibility to COVID-19: Focus in East Asian and European populations. *medRxiv*, 2020. DOI: [10.1101/2020.06.09.20126680](https://doi.org/10.1101/2020.06.09.20126680).
- (111) Bhattacharyya, C.; Das, C.; Ghosh, A.; Singh, A. K.; Mukherjee, S.; Majumder, P. P.; Basu, A.; Biswas, N. K. SARS-CoV-2 mutation 614G creates an elastase cleavage site enhancing its spread in high AAT-deficient regions. *Infect., Genet. Evol.* **2021**, *90*, 104760.
- (112) Shelton, J. F.; Shastri, A. J.; The 23andMe COVID-19 Team; Aslibekyan, S.; Auton, A. The UGT2A1/UGT2A2 locus is associated with COVID-19-related anosmia. *medRxiv*, 2021. DOI: [10.1101/2021.05.28.21257993](https://doi.org/10.1101/2021.05.28.21257993).
- (113) Neiers, F.; Jarriault, D.; Menetrier, F.; Briand, L.; Heydel, J. M. The odorant metabolizing enzyme UGT2A1: Immunolocalization and impact of the modulation of its activity on the olfactory response. *PLoS One* **2021**, *16* (3), No. e0249029.
- (114) Butowt, R.; von Bartheld, C. S. Anosmia in COVID-19: Underlying Mechanisms and Assessment of an Olfactory Route to Brain Infection. *Neuroscientist* **2020**, No. 107385842095690.
- (115) Torabi, A.; Mohammadbagheri, E.; Akbari Dilmaghani, N.; Bayat, A. H.; Fathi, M.; Vakili, K.; Alizadeh, R.; Rezaeimirghaed, O.; Hajiesmaeli, M.; Ramezani, M.; Simani, L.; Aliaghaei, A. Proinflammatory Cytokines in the Olfactory Mucosa Result in COVID-19 Induced Anosmia. *ACS Chem. Neurosci.* **2020**, *11* (13), 1909–1913.
- (116) Bryche, B.; St Albin, A.; Murri, S.; Lacôte, S.; Pulido, C.; Ar Gouilh, M.; Lesellier, S.; Servat, A.; Wasniewski, M.; Picard-Meyer, E.; Monchatre-Leroy, E.; Volmer, R.; Rampin, O.; Le Goffic, R.; Marianneau, P.; Meunier, N. Massive transient damage of the olfactory epithelium associated with infection of sustentacular cells by SARS-CoV-2 in golden Syrian hamsters. *Brain Behav Immun.* **2020**, *89*, 579–586.
- (117) Chen, M.; Shen, W.; Rowan, N. R.; Kulaga, H.; Hillel, A.; Ramanathan, M., Jr; Lane, A. P. Elevated ACE-2 expression in the olfactory neuroepithelium: implications for anosmia and upper respiratory SARS-CoV-2 entry and replication. *Eur. Respir. J.* **2020**, *56* (3), 2001948.
- (118) Cooper, K. W.; Brann, D. H.; Farruggia, M. C.; Bhutani, S.; Pellegrino, R.; Tsukahara, T.; Weinreb, C.; Joseph, P. V.; Larson, E. D.; Parma, V.; Albers, M. W.; Barlow, L. A.; Datta, S. R.; Di Pizio, A. COVID-19 and the Chemical Senses: Supporting Players Take Center Stage. *Neuron* **2020**, *107* (2), 219–233.
- (119) Klingenstein, M.; Klingenstein, S.; Neckel, P. H.; Mack, A. F.; Wagner, A. P.; Kleger, A.; Liebau, S.; Milazzo, A. Evidence of SARS-CoV2 Entry Protein ACE2 in the Human Nose and Olfactory Bulb. *Cells Tissues Organs* **2021**, *209* (4–6), 155–164.
- (120) Zugaj, M.; van Ditzhuijzen, N. S.; Golebski, K.; Fokkens, W. J. The effect of coronaviruses on olfaction: systematic review. *Rhinology* **2021**, *59* (3), 226–235.
- (121) Butowt, R.; Meunier, N.; Bryche, B.; von Bartheld, C. S. The olfactory nerve is not a likely route to brain infection in COVID-19: a critical review of data from humans and animal models. *Acta Neuropathol.* **2021**, *141* (6), 809–822.
- (122) de Melo, G. D.; Lazarini, F.; Levallois, S.; Hautefort, C.; Michel, V.; Larrous, F.; Verillaud, B.; Aparicio, C.; Wagner, S.; Gheusi, G.; Kerfoot, L.; Kornobis, E.; Donati, F.; Cokelaer, T.; Hervochon, R.; Madec, Y.; Roze, E.; Salmon, D.; Bourhy, H.; Lecuit, M.; Lledo, P. M. COVID-19-related anosmia is associated with viral persistence and inflammation in human olfactory epithelium and brain infection in hamsters. *Sci. Transl. Med.* **2021**, *13* (596), No. eabf8396.
- (123) Rodriguez, S.; Cao, L.; Rickenbacher, G. T.; Benz, E. G.; Magdamo, C.; Ramirez Gomez, L. A.; Holbrook, E.; Dhilla Albers, A.; Gallagher, R.; Westover, M. B.; Evans, K. E.; Tatar, D.; Mukerji, S.; Zafonte, R.; Boyer, E. W.; Yu, C. R.; Albers, M. W. Innate immune signaling in the olfactory epithelium reduces odorant receptor levels: modeling transient smell loss in COVID-19 patients. *medRxiv*, 2020. DOI: [10.1101/2020.06.14.20131128](https://doi.org/10.1101/2020.06.14.20131128).
- (124) Root-Bernstein, R. Anosmia-hyposmia and dysgeusia in COVID-19 may be due to SARS-CoV-2 protein mimicry of olfactory receptors. *Rhinology Online* **2020**, *3*, 148–151.
- (125) Walls, A. C.; Park, Y. J.; Tortorici, M. A.; Wall, A.; McGuire, A. T.; Veesler, D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell* **2020**, *181* (2), 281–292.
- (126) Johnson, B. A.; Xie, X.; Bailey, A. L.; Kalveram, B.; Lokugamage, K. G.; Muruato, A.; Zou, J.; Zhang, X.; Juelich, T.; Smith, J. K.; Zhang, L.; Bopp, N.; Schindewolf, C.; Vu, M.; Vanderheiden, A.; Winkler, E. S.; Swetnam, D.; Plante, J. A.; Aguilar, P.; Plante, K. S.; Popov, V.; Lee, B.; Weaver, S. C.; Suthar, M. S.; Routh, A. L.; Ren, P.; Ku, Z.; An, Z.; Debbink, K.; Diamond, M. S.; Shi, P. Y.; Freiberg, A. N.; Menachery, V. D. Loss of furin cleavage site attenuates SARS-CoV-2 pathogenesis. *Nature* **2021**, *591* (7849), 293–299.
- (127) Coutard, B.; Valle, C.; de Lamballerie, X.; Canard, B.; Seidah, N. G.; Decroly, E. The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. *Antiviral Res.* **2020**, *176*, 104742.
- (128) Cantuti-Castelvetri, L.; Ojha, R.; Pedro, L. D.; Djannatian, M.; Franz, J.; Kuivanen, S.; van der Meer, F.; Kallio, K.; Kaya, T.; Anastasina, M.; Smura, T.; Levanov, L.; Szirovicza, L.; Tobi, A.; Kallio-Kokko, H.; Österlund, P.; Joensuu, M.; Meunier, F. A.; Butcher, S. J.; Winkler, M. S.; Mollenhauer, B.; Helenius, A.; Gokce, O.; Teesalu, T.; Hepojoki, J.; Vapalahti, O.; Stadelmann, C.; Balistreri,

- G.; Simons, M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science* **2020**, *370* (6518), 856–860.
- (129) Daly, J. L.; Simonetti, B.; Klein, K.; Chen, K. E.; Williamson, M. K.; Antón-Plágaro, C.; Shoemark, D. K.; Simón-Gracia, L.; Bauer, M.; Hollandi, R.; Greber, U. F.; Horvath, P.; Sessions, R. B.; Helenius, A.; Hiscox, J. A.; Teesalu, T.; Matthews, D. A.; Davidson, A. D.; Collins, B. M.; Cullen, P. J.; Yamauchi, Y. Neuropilin-1 is a host factor for SARS-CoV-2 infection. *Science* **2020**, *370*, 861–865.
- (130) Jackson, B.; Boni, M. F.; Bull, M. J.; Colleran, A.; Colquhoun, R. M.; Darby, A. C.; Haldenby, S.; Hill, V.; Lucaci, A.; McCrone, J. T.; Nicholls, S. M.; O'Toole, A.; Pacchiarini, N.; Poplawski, R.; Scher, E.; Todd, F.; Webster, H. J.; Whitehead, M.; Wierzbicki, C.; Loman, N. J.; Connor, T. R.; Robertson, D. L.; Pybus, O. G.; Rambaut, A. Evaluating the Effects of SARS-CoV-2 Spike Mutation D614G on Transmissibility and Pathogenicity. *Cell* **2021**, *184* (1), 64–75.
- (131) Dao, T. L.; Hoang, V. T.; Colson, P.; Lagier, J. C.; Million, M.; Raoult, D.; Levasseur, A.; Gautret, P. SARS-CoV-2 Infectivity and Severity of COVID-19 According to SARS-CoV-2 Variants: Current Evidence. *J. Clin. Med.* **2021**, *10* (12), 2635.
- (132) Omotuyi, I. O.; Nash, O.; Ajiboye, O. B.; Iwegbulam, C. G.; Oyinloye, E. B.; Oyedele, O. A.; Kashim, Z. A.; Okaiyeto, K. Atomistic simulation reveals structural mechanisms underlying D614G spike glycoprotein-enhanced fitness in SARS-CoV-2. *J. Comput. Chem.* **2020**, *41* (24), 2158–2161.
- (133) Saha, P.; Majumder, R.; Chakraborty, S.; Kumar Srivastava, A.; Mandal, M.; Sarkar, S. Mutations in Spike Protein of SARS-CoV-2 Modulate Receptor Binding, Membrane Fusion and Immunogenicity: An Insight into Viral Tropism and Pathogenesis of COVID-19. *ChemRxiv*, 2020. DOI: [10.26434/chemrxiv.12320567.v1](https://doi.org/10.26434/chemrxiv.12320567.v1).
- (134) Jackson, C. B.; Zhang, L.; Farzan, M.; Choe, H. Functional importance of the D614G mutation in the SARS-CoV-2 spike protein. *Biochem. Biophys. Res. Commun.* **2021**, *538*, 108–115.
- (135) Hu, J.; He, C.-L.; Gao, Q.-Z.; Zhang, G.-J.; Cao, X.-X.; Long, Q.-X.; Deng, H.-J.; Huang, L.-Y.; Chen, J.; Wang, K.; Tang, N.; Huang, A.-L. D614G mutation of SARS-CoV-2 spike protein enhances viral infectivity. *bioRxiv*, 2020. DOI: [10.1101/2020.06.20.161323](https://doi.org/10.1101/2020.06.20.161323).
- (136) Tang, L.; Schulkens, A.; Chen, C.-N.; Deshayes, H.; Kenney, J. S. The SARS-CoV-2 Spike Protein D614G Mutation shows Increasing Dominance and May Confer a Structural Advantage to the Furin Cleavage Domain. *Preprints*, 2020. DOI: [10.20944/preprints202005.0407.v1](https://doi.org/10.20944/preprints202005.0407.v1).
- (137) Mansbach, R. A.; Chakraborty, S.; Nguyen, K.; Montefiori, D. C.; Korber, B.; Gnannakaran, S. The SARS-CoV-2 Spike variant D614G favors an open conformational state. *Sci. Adv.* **2021**, *7* (16), No. eabf3671.
- (138) Fernandez, A. Structural Impact of Mutation D614G in SARS-CoV-2 Spike Protein: Enhanced Infectivity and Therapeutic Opportunity. *ACS Med. Chem. Lett.* **2020**, *11* (9), 1667–1670.
- (139) Hou, Y. J.; Chiba, S.; Halfmann, P.; Ehre, C.; Kuroda, M.; Dinnon, K. H., 3rd; Leist, S. R.; Schäfer, A.; Nakajima, N.; Takahashi, K.; Lee, R. E.; Mascenik, T. M.; Graham, R.; Edwards, C. E.; Tse, L. V.; Okuda, K.; Markmann, A. J.; Bartelt, L.; de Silva, A.; Margolis, D. M.; Boucher, R. C.; Randell, S. H.; Suzuki, T.; Gralinski, L. E.; Kawaoka, Y.; Baric, R. S. SARS-CoV-2 D614G variant exhibits efficient replication ex vivo and transmission in vivo. *Science* **2020**, *370*, 1464–1468.
- (140) Ueha, R.; Kondo, K.; Kagoya, R.; Shichino, S.; Ueha, S.; Yamasoba, T. ACE2, TMPRSS2, and Furin expression in the nose and olfactory bulb in mice and humans. *Rhinology* **2020**, *59* (1), 105–109.
- (141) Shang, J.; Wan, Y.; Luo, C.; Ye, G.; Geng, Q.; Auerbach, A.; Li, F. Cell entry mechanisms of SARS-CoV-2. *Proc. Natl. Acad. Sci. U. S. A.* **2020**, *117* (21), 11727–11734.
- (142) Weissman, D.; Alameh, M. G.; de Silva, T.; Collini, P.; Hornsby, H.; Brown, R.; LaBranche, C. C.; Edwards, R. J.; Sutherland, L.; Santra, S.; Mansouri, K.; Gobeil, S.; McDanal, C.; Pardi, N.; Hengartner, N.; Lin, P. J. C.; Tam, Y.; Shaw, P. A.; Lewis, M. G.; Boesler, C.; Şahin, U.; Acharya, P.; Haynes, B. F.; Korber, B.; Montefiori, D. C. D614G Spike Mutation Increases SARS CoV-2 Susceptibility to Neutralization. *Cell Host Microbe* **2021**, *29* (1), 23–31.
- (143) Liu, Y.; Yan, L. M.; Wan, L.; Xiang, T. X.; Le, A.; Liu, J. M.; Periris, M.; Poon, L. L. M.; Zhang, W. Viral dynamics in mild and severe cases of COVID-19. *Lancet Infect. Dis.* **2020**, *20* (6), 656–657.
- (144) Hopkins, C.; Surda, P.; Whitehead, E.; Kumar, B. N. Early recovery following new onset anosmia during the COVID-19 pandemic - an observational cohort study. *J. Otolaryngol Head Neck Surg.* **2020**, *49* (1), 26.
- (145) Talavera, B.; García-Azorín, D.; Martínez-Pías, E.; Trigo, J.; Hernández-Pérez, I.; Valle-Peña Cob, G.; Simón-Campo, P.; de Lera, M.; Chavarria-Miranda, A.; López-Sanz, C.; Gutiérrez-Sánchez, M.; Martínez-Velasco, E.; Pedraza, M.; Sierra, Á.; Gómez-Vicente, B.; Guerrero, Á.; Arenillas, J. F. Anosmia is associated with lower in-hospital mortality in COVID-19. *J. Neurol. Sci.* **2020**, *419*, 117163.
- (146) Mascola, J. R.; Graham, B. S.; Fauci, A. S. SARS-CoV-2 Viral Variants-Tackling a Moving Target. *JAMA* **2021**, *325* (13), 1261–1262.
- (147) Hendy, M.; Kaufman, S.; Ponga, M. Molecular strategies for antibody binding and escape of SARS-CoV-2 and its mutations. *bioRxiv*, 2021. DOI: [10.1101/2021.03.04.433970](https://doi.org/10.1101/2021.03.04.433970).
- (148) Kemp, S. A.; Meng, B.; Ferriera, I. A. T. M.; Datir, R.; Harvey, W. T.; Papa, G.; Lytras, S.; Collier, D. A.; Mohamed, A.; Gallo, G.; Thakur, N.; The COVID-19 Genomics UK (COG-UK) Consortium; Carabelli, A. M.; Kenyon, J. C.; Lever, A. M.; De Marco, A.; Saliba, C.; Culap, K.; Cameroni, E.; Piccoli, L.; Corti, D.; James, L. C.; Bailey, D.; Robertson, D. L.; Gupta, R. K. Recurrent emergence and transmission of a sars-cov-2 spike deletion h69/v70. *bioRxiv*, 2021. DOI: [10.1101/2020.12.14.422555](https://doi.org/10.1101/2020.12.14.422555).
- (149) Graham, M. S.; Sudre, C. H.; May, A.; Antonelli, M.; Murray, B.; Varsavsky, T.; Kläser, K.; Canas, L. S.; Molteni, E.; Modat, M.; Drew, D. A.; Nguyen, L. H.; Polidori, L.; Selvachandran, S.; Hu, C.; Capdevila, J.; COVID-19 Genomics UK (COG-UK) Consortium; Hammers, A.; Chan, A. T.; Wolf, J.; Spector, T. D.; Steves, C. J.; Ourselin, S.; et al. Changes in symptomatology, reinfection, and transmissibility associated with the SARS-CoV-2 variant B.1.1.7: an ecological study. *Lancet Public Health* **2021**, *6* (5), e335–e345.
- (150) Mahase, E. Covid-19: Sore throat, fatigue, and myalgia are more common with new UK variant. *Brit Med. J.* **2021**, *372*, n288.
- (151) Office for National Statistics. *Covid-19 infection survey: Characteristics of people testing positive for covid-19 in England*, 2021. <https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/conditionsanddiseases/articles/coronaviruscovid19infectionsinthecommunityinengland/characteristicsofpeopletestingpositiveforcovid19inengland27january2021>.
- (152) Starr, T. N.; Greaney, A. J.; Hilton, S. K.; Ellis, D.; Crawford, K. H. D.; Dingens, A. S.; Navarro, M. J.; Bowen, J. E.; Tortorici, M. A.; Walls, A. C.; King, N. P.; Veesler, D.; Bloom, J. D. Deep Mutational Scanning of SARS-CoV-2 Receptor Binding Domain Reveals Constraints on Folding and ACE2 Binding. *Cell* **2020**, *182* (5), 1295–1310.
- (153) Mak, G. C. K.; Lau, A. W. L.; Chan, A. M. Y.; Lam, E. T. K.; Chan, R. C. W.; Tsang, D. N. C. The surveillance of spike protein for patients with COVID-19 detected in Hong Kong in 2020. *J. Med. Virol.* **2021**, *93* (9), 5644–5647.
- (154) Bidkar, V.; Mishra, M.; Selvaraj, K.; Joshi, P.; H, S. B.; Dabhekar, S.; Prathipati, K. K.; Rathod, B. S.; Shendre, P.; Gondode, P. Testing Olfactory and Gustatory Dysfunctions among Quarantine COVID-19 Suspects. *Indian J. Otolaryngol Head Neck Surg.* **2021**, *73* (3), 304–309.
- (155) Dravid, A. N.; Mane, D. N.; Khan, Z. A. Neurological issues during severe COVID-19 in a tertiary level hospital in Western India. *Neurosci. Lett.* **2021**, *749*, 135692.
- (156) Bhatta, S.; Sharma, D.; Sharma, S.; Maharjan, L.; Bhattachan, S.; Shah, M. K.; Singhal, A.; Ghanpur, A. D.; Ganesuni, D.; Saindani, S. J. Smell and Taste Disturbance in COVID-19 Patients: A Prospective Multicenteric Review. *Indian J. Otolaryngol Head Neck Surg.* **2021**, *1*–7.

- (157) Patil, K. I.; Prabhu, R. M.; Jose, P.; Chandrashekappa, H.; Viswanatha, B. Anosmia and ageusia in Covid-19 patients. *Sch J Otolaryngol* **2020**, *5* (3), 485–489.
- (158) DerSimonian, R.; Laird, N. Meta-analysis in clinical trials. *Controlled Clin. Trials* **1986**, *7* (3), 177–188.
- (159) Higgins, J. P.; Thompson, S. G. Quantifying heterogeneity in a meta-analysis. *Stat Med* **2002**, *21* (11), 1539–1558.
- (160) Egger, M.; Smith, G. D.; Phillips, A. N. Meta-analysis: principles and procedures. *Brit Med. J.* **1997**, *315* (7121), 1533–1537.